



## Germination and Growth Parameters in Sorghum Cultivars (*Sorghum bicolor* L.) Effected by Boron Application Under Salinity Stress

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### ABSTRACT

The aim of this research is investigating the effects of boron on the germination and growth of three sorghum cultivars at different salinity stress levels. The experiment was arranged as four replications according to the factorial experimental design in completely random blocks. The three sorghum (*Sorghum bicolor* L.) cultivars (Erdurmuş, Uzun and Gözde 80) selected for the genetic material. NaCl compound was utilized as salt source and solutions were prepared at concentrations of 0-75-150 mM. Boron was applied as H<sub>3</sub>BO<sub>3</sub> at 0-5-10-15 mM. In general regarding growth parameters, the values obtained in Gözde 80 cultivar were determined as the highest averages. Whereas the salinity levels effect was examined in this study, a decrease was determined in the parameters measured as the level of the stress factor increased. Salinity had a high adverse effect at the 150 mM level, and as expected the highest averages were obtained in the control treatments. Low-dose boron applications have positive effects on germination and growth parameters in this experiment. Therewithal under salinity stress conditions, low-dose boron applications showed affirmative efficacy compared to the control of each condition. In this experiment, determined that boron applications reduce this effect under salinity stress conditions that sorghum seeds may encounter during the germination period, but the boron dose level to be applied should be properly controlled.

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## Tuzluluk Stresi Altında Bor Uygulamasından Etkilenen Sorgum Çeşitlerinde (*Sorghum bicolor* L.) Çimlenme ve Büyüme Parametreleri

### ÖZET

Bu araştırmanın amacı, farklı tuz stresi seviyelerinde üç sorgum çeşidinin çimlenmesi ve büyümesi üzerine borun etkilerini araştırmaktır. Deneme, tesadüf blokları faktöriyel deneme deseni kullanılarak dört tekerrürlü olarak düzenlenmiştir. Genetik materyal için seçilen üç sorgum çeşidi (*Sorghum bicolor* L.) Erdurmuş, Uzun ve Gözde 80'dir. Tuz kaynağı olan NaCl bileşiğinden 0-75-150 mM konsantrasyonlarda çözeltiler hazırlanarak stres koşulları oluşturulmuştur. Bor dozları ise 0-5-10-15 mM olarak H<sub>3</sub>BO<sub>3</sub> kimyasalından hazırlanarak uygulanmıştır. Genel olarak Gözde 80 çeşidinde elde edilen değerler en yüksek ortalamalar olarak belirlenmiştir. Bu çalışmada tuzluluk düzeylerinin etkisi incelendiğinde, stres faktörünün düzeyi arttıkça ölçülen parametrelerde azalmalar tespit edilmiştir. En düşük ortalamalar 150 mM seviyesinde tuz uygulamalarında elde edilirken, en yüksek ortalamalar kontrol uygulamasında belirlenmiştir. Düşük dozlarda bor uygulamaları bu çalışmada tüm parametreler üzerinde artışlar meydana getirmiştir. Tuzluluk stresi koşullarında düşük dozda bor uygulamaları her koşulun kontrolüne göre pozitif etki göstermiştir. Bu denemede, sorgum tohumlarının çimlenme döneminde karşılaşılabilecekleri tuzluluk stresi koşullarında bor uygulamalarının bu etkiyi azalttığı ancak uygulanacak bor doz seviyesinin uygun şekilde kontrol edilmesi gerektiği belirlenmiştir.

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## INTRODUCTION

The request for food is spawning day by day with the rapid increase in the world population. However, human activities and geological processes cause adverse biotic or abiotic agents, especially salinity problems (Kumar et al., 2017; Guo et al., 2022). Salinity is a major hazard in arid and semi-arid climatic areas and is a significant stunting factor in universal food manufacture (Ahmed et al., 2020; Tolay, 2021). Salinity occurs when soluble salts that accumulate in groundwater by washing in arid and semi-arid climatic regions rise to the soil surface with capillarity due to high groundwater and accumulate on the surface as a result of water evaporation. Salinity stress primarily occurs in degradations in metabolic and functional mechanisms. Than, it causes injury to plant organs, reduce in product quality and even death (Shabala, 2009; Arslan et al., 2013).

Salinity is one of main abiotic stress limiting plant growth, development and productivity. The significant inhibitory effect of salinity on plant growth and crop is due to (1) osmotic effect, (2) ion toxicity, and (3) nutritional deficiency leading to reduced photosynthetic effect and other physiological disorder (Almodares et al, 2008).

Nowadays, it is stated that there is a substantial decrease in crop yield due to global soil salinization and salt effect is approximately millions hectares (Pirasteh-Anosheh et al., 2017; Asgari & Diyanat, 2021; Karle et al., 2021). Therefore, it has turned into immediate needs to increase plants grown in salty areas and utilized for food and treatment and to discover a solution to salinity stress (Shahid et al., 2021).

Sorghum (*Sorghum bicolor* L.) is member of a cereal crop family with various utilizes food, forage crop (Baran & Kocabağlı, 2000; Arslan et al., 2017) and energy (Rooney & Waniska, 2000; Ribas, 2014). Whereat it is from the cereal family, it can be easily made into silage (Vadakkekara Joseph, 2016; Arslan et al., 2017) and as a result, it can be fed to animals (Karadağ & Özkurt 2014). Sorghum, studied annually (Pontieri et al., 2012) by breeders, has wide genetic changeability (Tigabu et al., 2012; Shakeri et al., 2017) and germplasm origins to adopt new varieties into multiple ecoregions (Bibi et al., 2016). Sorghum is one of the C4 plants that is moderately saline tolerant (Maas & Hoffman, 1986) and highly adaptable to semi-arid and arid regions where salinity creates problems for vegetative production (Kebede et al., 2001; Ejeta & Knoll, 2007; Ali et al., 2011).

Boron has a significant role in physiological courses within plants and is therefore one of the most major micronutrients for their growth (Warrington, 1923). This is one of the most limiting nutrient for grains especially for lucubratory varieties growing in sandy soils where it can vanish by filtering due to its high mobility in the soil (Uraguchi & Fujiwara, 2011). There are differences in boron access in plant species and varieties (Hu & Brown, 1997). Within its roles, the element boron is associated with increasing seed yield and quality (Batista et al., 2021). Excessive boron intake limits plant growth and damages the photosynthetic system. Thereby causing cirtical physiological reactions such as degradation of auxin biosynthesis leading to decreased fruit or crop yields. Since boron is an element involved in cell growth, nucleic acid formation, nitrogen fixation, cell wall biosynthesis, sugar translocation, as well as flowering and pollen tube growth, its importance is further increased under stress conditions (Marschner, 2012).

It is obtained that germination and emergence processes in sorghum breeding are the most enlightening phases of the plant life cycle to determine the effect of salinity (Krishnamurthy et al., 2017). It is necessary to develop new strategies and appropriate methods in order to reduce the negative effects of low salt stress that plants are exposed to during the germination and seedling stages of sorghum cultivation. In these cases, it has been found that the seed can stimulate some metabolic processes in germination by pretreatment with some plant growth regulators and increase seed emergence under varied environmental conditions (Ren et al., 2020). Therefore, we hypothesized that the adverse effects at different salt stress levels could be mitigated by boron application. The aim of the experiment was to examine the probability of differential salt stress reduction at the seedling stage of three sorghum varieties by applying boron levels.

## MATERIAL ve METHOD

As investigating the effects of boron on the germination and seedling growth of three sorghum cultivars at different salt stress levels, this experiment was carried out in the Forage Crops Laboratory of the Field Crops Department of Akdeniz University. The trial was arranged in four replications using factorial arrangement according to the completely randomized blocks experimental design. The three sorghum cultivars (*Sorghum bicolor* L.) selected for the genetic material are Erdurmuş, Uzun and Gözde 80 and were

obtained from the Western Mediterranean Agricultural Research Institute.

Petri dishes were rinsed with deionized water, sterilized and incubated as a general protocol procedure. Twenty sorghum seeds in same size of each cultivar were placed evenly in a petri dish (9 cm diameter) using forceps after two sterilized Whatman Filter papers. NaCl (Sigma®) compound was utilized as salt source and solutions were prepared at concentrations of 0-75 mM-150 mM. Boron was applied as H<sub>3</sub>BO<sub>3</sub> at 0-5-10-15 mM. To prevent evaporation, all the petri dishes were wrapped with parafilm. The petri dishes were settled in a growth chamber at 20°C under photoperiodic condition 16 hours light 8 hours dark. Observations were recorded daily. The study ended on the eleventh day.

Germination tests were carried out according to ISTA rules (2017). Radicle and plumula lengths of seedling were measured using a scale. The seed of germination (MGT) was calculated using formulas described by Majda et al. (2019). Germination rate (GR) was calculated according to Xia et al. (2019). Germination index (GI) and seedling vigor index (SVI) were counted by the method of Xia et al. (2019). The radicle/plumula ratio (R/S ratio) was calculated as the following equation (Shtaya et al., 2021). The calculation of stress tolerance indices formulas as described by Naraw et al. (2014).

$$\text{MGT (day): } \frac{\sum \text{number of seeds germinated on the } i^{\text{th}} \text{ day}}{\text{number of days to count the } n^{\text{th}}}$$

GR(%):number of germinated seed/total number of seed tested\*100

$$\text{GI: } \frac{\sum \text{the number of germinated seeds in day}}{\text{day of counting seed germination}}$$

SVI: germination percentage\*average seedling length/100

R/P ratio: Radicle length/ plumula length

SLSI (%): (Plumula length of stressed plant/ Plumula length of control)\*100

RLSI (%): (Radicle length of stressed plant/ Radicle length of control)\*100

SFSI (%): (Plumula fresh weight of stressed plant/ Plumula fresh weight of control)\*100

RFSI (%): (Radicle fresh weight stress of stressed plant/ Radicle fresh weight of control)\*100

### Statistic Analysis

Datas determined for the study subjected to analysis variance using R (ANOVA) and compared with Duncan's multiple range, which differed significantly at 0.05 levels. (4.3.19)

### RESULTS and DISCUSSION

The variance analysis of the different boron applications on germination and seedling parameters of sorghum cultivars affected by salinity conditions are given in Table 1 and Table 2.

Table 1. Results of variance analysis on growth parameters of boron doses and salinity in sorghum cultivars  
*Çizelge 1. Tuz stresi seviyelerinde üç sorgum çeşidinin büyüme parametreleri üzerine borun etkileri*

Source of variance	df	PFW	RFW	TB	PLSI	RLSI	PFSI	RFSI
C	2	21.90**	20.26**	0.20	615.00**	75.00**	227.00**	1655.00**
S	2	3529.70**	319.92**	5661.70**	83691.00**	74442.00**	84182.00**	77026.00**
B	3	4646.30**	235.67**	7337.50**	948.00**	779.00**	701.00**	510.00**
C*S	4	29.80**	26.68**	62.50**	184.00**	255.00**	78.00*	905.00**
C*B	6	23.50**	12.83**	49.60**	187.00**	124.00**	41.00	338.00**
S*B	6	213.50**	10.59**	349.00**	253.00**	211.00**	192.00**	180.00**
C*S*B	12	6.90**	7.46**	11.70**	104.00**	82.00**	14.00	146.00**

\*Significant at the 0.05 probability level. \*\*Significant at the 0.01 probability level.

(Cultivar (Çeşit): C, Salinity (Tuzluluk): S, Boron (Bor): B, Mean germination time (Ortalama Çimlenme Zamanı): MGT, Germination Rate (Çimlenme oranı): GR, Germination index (Çimlenme indeksi): GI, Seedling Vigor index (Fide canlılık indeksi): SVI, Plumula length (Plumula uzunluğu): PL, Radicle length (Radicle uzunluğu): RL, Radicle/Plumula rate (Radicle/ plumula oranı): R/S)

According to analysis of variance, all parameters except total biomass were statistically significant at the level of 1% in all cultivars. Similarly, increasing salinity levels, boron doses and their interactions had a significant effect (p<0.01) on the germination and seedling parameters of sorghum cultivars. Closely, the interaction of cultivars-boron doses and full interaction factor showed meanful effects (p<0.01) on all parameters except plumula fresh weight stress tolerance index. The plumula fresh weight stress

tolerance index was effected at 5% levels, as the other germination and seedling parameters were significantly (p<0.01) influenced by the interaction between cultivars and salinity levels (Table 1 and 2).

In general, the values obtained in Gözde 80 cultivar were determined as the maximum averages. The specified parameters are MGT, PL, RL, R/P rate and RFW. However, the highest germination rate and seedling weight were found in Erdumuş cultivar (Table 3a-3b).

Table 2. Results of variance analysis on growth parameters and stress tolerances of boron doses and salinity in sorghum cultivars

*Çizelge 2. Tuz stresi seviyelerinde üç sorgum çeşidinin büyüme parametreleri ve stres toleransları üzerine borun etkileri*

Source of variance	df	PFW	RFW	TB	PLSI	RLSI	PFSI	RFSI
C	2	21.90**	20.26**	0.20	615.00**	75.00**	227.00**	1655.00**
S	2	3529.70**	319.92**	5661.70**	83691.00**	74442.00**	84182.00**	77026.00**
B	3	4646.30**	235.67**	7337.50**	948.00**	779.00**	701.00**	510.00**
C*S	4	29.80**	26.68**	62.50**	184.00**	255.00**	78.00*	905.00**
C*B	6	23.50**	12.83**	49.60**	187.00**	124.00**	41.00	338.00**
S*B	6	213.50**	10.59**	349.00**	253.00**	211.00**	192.00**	180.00**
C*S*B	12	6.90**	7.46**	11.70**	104.00**	82.00**	14.00	146.00**

\*Significant at the 0.05 probability level. \*\*Significant at the 0.01 probability level.

Table 3a. The effects of boron doses on growth parameters of sorghum cultivars exposed to salinity

*Çizelge 3a. Farklı tuz stresi seviyelerine maruz kalmış üç sorgum çeşidinin büyüme parametreleri üzerine borun etkileri*

C	MGT (day)	GR (%)	GI (%)	SVI (%)	PL (cm)	RL (cm)
E	3.91±0.34 <sup>b</sup>	89.58±8.86 <sup>a</sup>	91.88±9.09 <sup>b</sup>	48.55±4.80 <sup>c</sup>	5.42±1.91 <sup>b</sup>	4.75±1.99 <sup>c</sup>
U	4.04±0.24 <sup>a</sup>	79.17±14.06 <sup>b</sup>	99.58±14.81 <sup>a</sup>	52.88±9.03 <sup>a</sup>	5.31±1.94 <sup>c</sup>	5.09±2.92 <sup>b</sup>
G	3.76±0.37 <sup>c</sup>	79.80±13.81 <sup>b</sup>	83.99±17.20 <sup>c</sup>	51.21±9.13 <sup>b</sup>	6.10±2.30 <sup>a</sup>	6.16±2.34 <sup>a</sup>

Different letters next to values indicate statistically different means at p<0.05 level, and p<0.01 levels.

*Ortalamaları temsil eden farklı harfler, p<0.05 düzeyinde ve p<0.01 düzeyinde istatistiksel olarak farklı ortalamaları gösterir.*

Table 3b. The effects of boron doses on growth parameters of sorghum cultivars exposed to salinity

*Çizelge 3b. Farklı tuz stresi seviyelerine maruz kalmış üç sorgum çeşidinin büyüme parametreleri üzerine borun etkileri*

C	R/P rate	PFW (mg)	RFW (mg)	TB (mg)
E	0.85±0.09 <sup>b</sup>	35.99±11.80 <sup>a</sup>	6.63±2.16 <sup>c</sup>	42.62±13.91
U	0.95±0.19 <sup>a</sup>	35.00±11.82 <sup>b</sup>	7.51±4.12 <sup>b</sup>	42.51±15.86
G	0.96±0.09 <sup>a</sup>	34.70±14.58 <sup>b</sup>	7.90±3.89 <sup>a</sup>	42.57±18.52

Different letters next to values indicate statistically different means at p<0.05 level, and p<0.01 levels.

*Ortalamaları temsil eden farklı harfler, p<0.05 düzeyinde ve p<0.01 düzeyinde istatistiksel olarak farklı ortalamaları gösterir.*

As there was no statistical difference between the total biomass, the averages were determined as nearly 43.00 mg in cultivars (Table 3b). Significant differences in biomass were not only due to genetic makeup but also to the growth medium (Amano et al., 1996). The reason why a difference was not obtained in this study is thought to be due to the fact that it was made in a petri dish. Mean germination time, is one of the most distinctive features of the cultivars, was obtained in Erdurmuş, Uzun and Gözde-80 cultivars as 3.91 days, 4.04 days and 3.76 days, respectively (Table 3a).

In this experiment, it was concluded that different morphological features showed different responses to applications. In addition, it was observed that the applications helped the tolerance under stress conditions on the cultivars. In most cases, germination traits were useful in predicting the suitability of seeds for commercial usage due to their quality at the seedling stage, whereas germination rate affects seed emergence uniformity (Ching, 1959; Thomson & El-Kassaby, 1993; El-Kassaby, 2000, El-Kassaby et al., 2008).

As the effect of salinity levels was examined in this

study, a decrease was determined in the parameters measured as the level of the stress factor increased. The lowest averages were obtained in salt applications at the level of 150 mM, and the highest values were determined in the control (Table 4a,4b). A high salt concentration was a limiting agent for initiating and maintaining germination, decreasing the amount of water available in the presence of salt and this affects both germination percentage and germination speed (dos Santos et al., 2019; Moghaddam et al., 2020).

In this study and many experiment, as the NaCl concentration increased, the seeds generally had lower germination than the control (Table 4a, Atta et al., 2021); however, it was observed that the mean germination day increased (Table 4a, Kara et al., 2011; Li et al., 2016; Önal Aşçı & Üney, 2016) and germination index (Table 4a, Carpıcı et al., 2009; Aishah et al., 2010; Atış, 2011).

As the salt stress level increased, 23.44% and 37.50% decrease in plumula length and 25.86% and 45.57% reduction in radicle length were determined, respectively (Table 4a). The obtained decreases were found to be compatible with other studies. (Rajakumar,

2013). In terms of salt stress tolerance, root and stem lengths are significant parameters (Arslan & Aydinoğlu, 2018), whereat nutrients and water are get into the plant through direct contact with the roots and subscribe to shoot development (Haileselassie & Gselassie, 2012). Similar to these results, reductions of 26.68% and 38.54% in plumula fresh weight and 32.54% and 50.19% in radicle fresh weight,

respectively, were observed. In other words, as can be seen from these results, 150 mM salt application caused a half reduction in radicle fresh weight (Table 4b, Datta et al., 2009; Yildirim et al., 2011; Ödemiş et al., 2019; Bhati et al., 2021; Tenikecier & Ates, 2022; Quamruzzaman et al., 2022). Thus, it was determined that the radicle was more precision to salinity than the plumula (Kaya et al., 2003).

Table 4a. The growth parameters of sorghum cultivars exposed to salinity

*Çizelge 4a. Sorgum çeşitlerinde tuzluluğun büyüme parametreli üzerine etkisi*

SL (mM)	MGT (day)	GR (%)	GI (%)	SVI (%)	PL (cm)	RL (cm)
0 mM	3.68±0.37 <sup>c</sup>	90.94±10.55 <sup>a</sup>	100.67±13.57 <sup>a</sup>	55.80±6.97 <sup>a</sup>	7.04±2.26 <sup>a</sup>	7.00±2.81 <sup>a</sup>
75 mM	3.90±0.28 <sup>b</sup>	83.13±11.04 <sup>b</sup>	92.26±12.87 <sup>b</sup>	51.13±6.60 <sup>b</sup>	5.39±1.60 <sup>b</sup>	5.19±2.07 <sup>b</sup>
150 mM	4.13±0.22 <sup>a</sup>	74.48±12.85 <sup>c</sup>	82.52±14.26 <sup>c</sup>	45.71±7.31 <sup>c</sup>	4.40±1.34 <sup>c</sup>	3.81±1.27 <sup>c</sup>

Different letters next to values indicate statistically different means at p<0.05 level, and p<0.01 levels.

*Ortalamaları temsil eden farklı harfler, p<0.05 düzeyinde ve p<0.01 düzeyinde istatistiksel olarak farklı ortalamaları gösterir.*

Table 4b. The growth parameters of sorghum cultivars exposed to salinity

*Çizelge 4b. Sorgum çeşitlerinde tuzluluğun büyüme parametreli üzerine etkisi*

SL (mM)	R/P rate	PFW (mg)	RFW (mg)	TB (mg)
0 mM	0.97±0.13 <sup>a</sup>	44.27±14.30 <sup>a</sup>	10.14±3.61 <sup>a</sup>	53.91±18.03 <sup>a</sup>
75 mM	0.94±0.16 <sup>b</sup>	34.23±7.19 <sup>b</sup>	6.84±2.90 <sup>b</sup>	41.56±11.63 <sup>b</sup>
150 mM	0.86±0.11 <sup>c</sup>	27.21±9.29 <sup>c</sup>	5.05±1.62 <sup>c</sup>	32.26±8.73 <sup>c</sup>

Different letters next to values indicate statistically different means at p<0.05 level, and p<0.01 levels.

*Ortalamaları temsil eden farklı harfler, p<0.05 düzeyinde ve p<0.01 düzeyinde istatistiksel olarak farklı ortalamaları gösterir.*

The salinity condition reduces the total photosynthetic capacity of the cultivars and the amount of chlorophyll level, thus reducing the synthesis of assimilate required for plant growth, and as a result, it is now clear that it causes a shrinkage in plant biomass (Taiz et al., 2015; Pirasteh-Anosheh & Hedayati-Firoozabadi, 2022). Another reason for the loss of

biomass may be the decrease in the metabolic energy of the plant due to adaptation, the photosynthesis rate of the leaf, the decrease in carbon uptake, and tissue damage if the plant reaches the maximum salt concentration that the plant can tolerate (Pirasteh-Anosheh et al., 2016).

Table 5a. The effects of boron doses on growth parameters of sorghum cultivars

*Çizelge 5a. Bor uygulamalarının sorgum çeşitlerin büyüme parametreleri üzerine etkileri*

Boron (mM)	MGT (day)	GR (%)	GI (%)	SVI (%)	PL (cm)	RL (cm)
0 Mm	3.91±0.28 <sup>c</sup>	92.78±8.14 <sup>a</sup>	97.11±10.35 <sup>b</sup>	53.84±5.07 <sup>b</sup>	6.54±1.11 <sup>b</sup>	6.39±1.81 <sup>b</sup>
5 Mm	3.51±0.28 <sup>d</sup>	87.92±6.37 <sup>b</sup>	103.15±10.39 <sup>a</sup>	57.24±5.53 <sup>a</sup>	7.28±1.77 <sup>a</sup>	7.31±2.24 <sup>a</sup>
10 Mm	4.03±0.24 <sup>b</sup>	83.61±8.83 <sup>c</sup>	92.78±10.55 <sup>c</sup>	51.42±5.00 <sup>c</sup>	5.58±1.51 <sup>c</sup>	5.28±1.66 <sup>c</sup>
15 Mm	4.17±0.13 <sup>a</sup>	67.08±12.33 <sup>d</sup>	74.23± 12.74 <sup>d</sup>	41.01±5.74 <sup>d</sup>	3.04±0.62 <sup>d</sup>	2.35±0.44 <sup>d</sup>

Different letters next to values indicate statistically different means at p<0.05 level, and p<0.01 levels.

*Ortalamaları temsil eden farklı harfler, p<0.05 düzeyinde ve p<0.01 düzeyinde istatistiksel olarak farklı ortalamaları gösterir.*

Table 5b. The effects of boron doses on growth parameters of sorghum cultivars

*Çizelge 5b. Bor uygulamalarının sorgum çeşitlerin büyüme parametreleri üzerine etkileri*

Boron (mM)	R/P rate	PFW (mg)	RFW (mg)	TB (mg)
0 Mm	0.96±0.13 <sup>b</sup>	39.30±8.68 <sup>b</sup>	8.00±2.84 <sup>b</sup>	47.98±10.97 <sup>b</sup>
5 Mm	1.00±0.12 <sup>a</sup>	47.30±11.31 <sup>a</sup>	10.34±3.48 <sup>a</sup>	57.64±14.44 <sup>a</sup>
10 Mm	0.94±0.11 <sup>b</sup>	34.01±6.83 <sup>c</sup>	6.86±2.24 <sup>c</sup>	40.87±8.74 <sup>c</sup>
15 Mm	0.78±0.08 <sup>c</sup>	20.29±3.65 <sup>d</sup>	4.18±2.29 <sup>d</sup>	23.80±4.40 <sup>d</sup>

Different letters next to values indicate statistically different means at p<0.05 level, and p<0.01 levels.

*Ortalamaları temsil eden farklı harfler, p<0.05 düzeyinde ve p<0.01 düzeyinde istatistiksel olarak farklı ortalamaları gösterir.*

Low-dose boron applications increased all parameters in this study and Alamri et al. (2018) showed an increase only on the germination day, as there was an

inverse relationship in this parameter only on the germination day. High boron doses had a negative effect on both plant shoot and root length (Table 5a-5b). Among the functions of boron element due to its

functions in plant metabolism, it is related to increasing seed yield and quality (Silva-Matos et al., 2017; Batista et al., 2021). The fact that high boron levels induce harmful effects on plant growth (Bhamburdekar, 2002) and development (Tariq & Mott, 2006) is remarkable (Jadhav & Bhamburdekar, 2014). In addition, excess boron primarily inhibits plant germination and cell division and damages the thylakoid assembly by affecting photosynthesis, thereby reducing CO<sub>2</sub> absorption, resulting in reduced

root and shoot growth (Reguera et al., 2009; Uluisik et al., 2018).

In general, it was determined that the effect of salinity on the growth of different cultivars decreased linearly. Considering the cultivars, the plumule length decreased by 19.00% and 35.44% in Erdurmuş, by 32.68% and 42.82% in Uzun, and by 18.40% and 33.96% in Gözde-80 (Figure1).

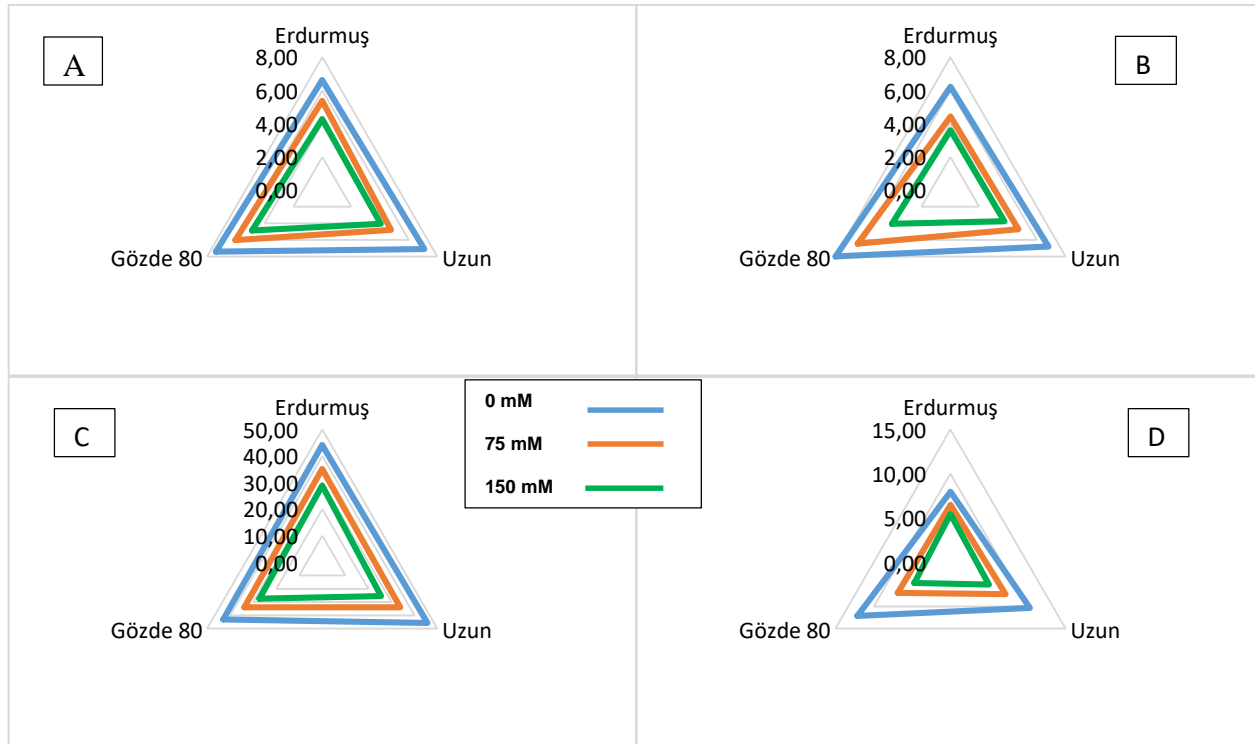


Figure 1. The effects of salinity on a. plumula length (cm), b. radicle length (cm), c. plumula fresh weight (mg) and d. radicle fresh weight (mg) in sorghum cultivars

Şekil 1. Sorgum çeşitlerinde tuzluluğun a. plumula uzunluğunda (cm), b. kök uzunluğunda (cm), c. plumula taze ağırlığında (mg) ve d. kök taze ağırlığında (mg) etkileri

On the other hand, in the same cultivars salinity stress decreased the radicle length by 28.94%, 42.12%, 30.84%, 44.94%, 19.20% and 48.94%, respectively. The highest plumule fresh weight was found in the control application and the lowest was in the application of 150 mM salt at Uzun cultivar. At the same time, when the radicle was examined in terms of fresh weight, the highest average was determined in the control application (12.12 mg) in Gözde80 cultivar and the lowest average was determined in the 150 mM application (4.70 mg) in the same variety. As a result of the reductions, it was found that Uzun is the most sensitive variety to salinity in general (Figure 1).

Whenever the effects of increasing boron doses on the cultivars were examined, the highest averages in length parameters were obtained at a dose of 5 mM in Gözde 80 cultivars, and the lowest averages were determined at a dose of 15 mM in Erdurmuş cultivars.

However, when looking at the fresh weights, the highest and lowest means were obtained in Uzun cultivar at 5 mM and 15 mM doses, respectively (Figure 2). Low boron dose has a positive effect on plant growth up to 5 mM dose, which can be explained by the effectiveness of boron element in plants such as cell division, elongation in meristematic tissues, membrane integrity, cell wall formation, leaf expansion (Marschner, 2012; Gupta & Solanki, 2013; Da Rocha Pinho et al., 2015).

## CONCLUSION

As a result of this study, different salinity levels created with NaCl decreased germination and growth parameters. Differences and responses between cultivars were also observed. In addition, determined that low-level boron applications caused an increase in the parameters examined under increasing NaCl

conditions, yet observed that high-level applications caused a decrease in the parameters. It is thought that the effects of different salinity levels and boron

applications determined in this study on germination and growth will be beneficial for further studies.

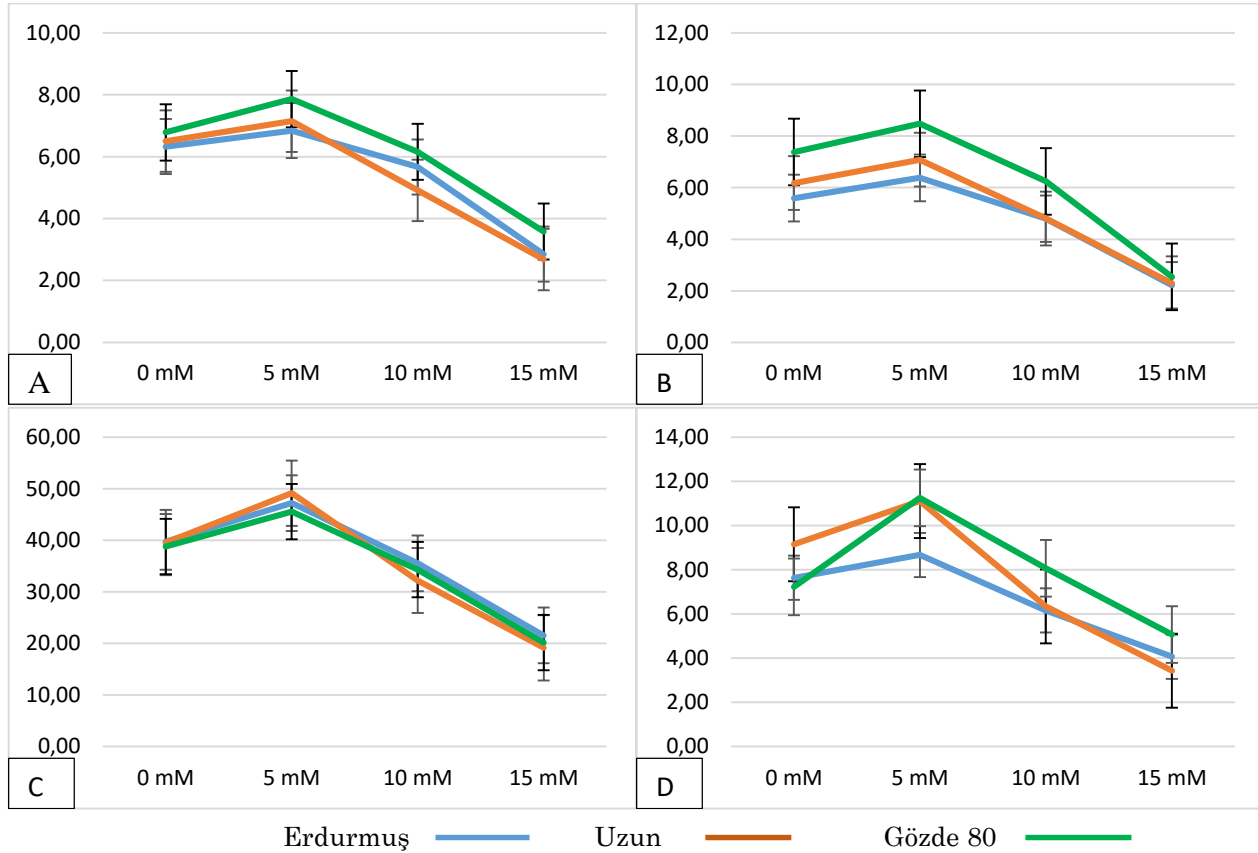


Figure 2. The effects of boron doses on a. plumula length (cm), b. radicle length (cm), c. plumula fresh weight (mg) and d. radicle fresh weight (mg) in sorghum cultivars

Şekil 2. Sorghum çeşitlerinde bor uygulamalarının a. plumula uzunluğunda (cm), b. kök uzunluğunda (cm), c. plumula taze ağırlığında (mg) ve d. kök taze ağırlığında (mg) etkileri

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#### Researchers Contribution Rate Declaration Summary

The authors declared that they have contributed to the article equally. All authors discussed the results and contributed to the final manuscript.

#### Statement of Conflict of Interest

Author has declared no conflict of interest.

#### REFERENCES

Ahmed, K., Qadir, G., Nawaz, M.Q., Riaz, M.A., Nawaz, M.F., & Ullah, M.M.A. (2020). Combined effect of growth hormones and gypsum induces salinity tolerance in wheat under saline-sodic soil. *Journal of Animal and Plant Sciences*, 31(1), 121–130. <https://doi.org/10.36899/JAPS.2021.1.0200>.

Aishah, H.S., Saberi, A.R., Halim, R.A., & Zaharah, A.R. (2010). Salinity effects on germination of forage sorghumes. *Journal of Agronomy*, 9(4), 169–174.

Alamri, S.A., Siddiqui, M.H., Al-Khaishani, M.Y., & Hayssam, M.A. (2018). Boron induces seed germination and seedling growth of *Hordeum vulgare* L. under NaCl stress. *Journal of Advances in Agriculture*, 8(1), 1224-1234.

Ali, M.A., Abbas, A., Awan, S.I., Jabran, K., & Gardezi, S.D.A. (2011). Correlated response of various morphophysiological characters with grain yield in sorghum landraces at different growth phases. *Journal Animal Plant Science*, 21, 671-679.

Almodares, A., Hadi, M.R., & Dosti, B. (2008). The effects of salt stress on growth parameters and carbohydrates contents in sweet sorghum. *Res J. Environ Sci*, 2(4), 298-304.

Amano, T., Shi, C.J., Qin, D.L., Tsuda, M., & Matsumoto, Y. (1996). High-Yielding Performance of Paddy Rice Achieved in Yunnan Province, China: I. High yielding ability of Japonica F1 hybrid rice, Yu-Za 29. *Japan J. Crop Sci.*, 65(1), 16–21.

- Arslan, M., Erdurmuş, C., & Çakmakçı, S. (2013). Effects of NaCl concentrations on germination and early seedling growth of silage sorghum (*Sorghum bicolor* (L.) Moench) varieties on different textured soils. *Journal of Food, Agriculture & Environment*, 11(2), 474-476.
- Arslan, M., Erdurmuş, C., Öten, M., Aydınoglu, B., & Çakmakçı, S. (2017). Sorgum ve bazı bitkilerin ile farklı oranlarda karışımlarından hazırlanan silajların kalite özellikleri. *Tekirdağ Ziraat Fakültesi Dergisi*, 14, 34-41.
- Arslan, M. & Aydınoglu, B. (2018). Tuzluluk (NaCl) stresinin mürdümükde (*Lathyrus sativus* L.) çimlenme ve erken fide gelişme özelliklerine etkisi. *Akademik Ziraat Dergisi*, 7(1), 49-54.
- Asgari, F. & Diyanat, M. (2021). Effects of silicon on some morphological and physiological traits of rose (*Rosa chinensis* var. minima) plants grown under salinity stress. *Journal of Plant Nutrition*, 44(4), 536-549. <https://doi.org/10.1080/01904167.2020.1845367>
- Atış, İ. (2011). Bazı silajlık sorgum (*Sorghum bicolor* L. Moench) çeşitlerinin çimlenmesi ve fide gelişimi üzerine tuz stresinin etkileri. *Süleyman Demirel Üniversitesi Ziraat Fakültesi Dergisi*, 6(2), 58-67.
- Atta, K., Pal, A.K., & Jana, K. (2021). Effects of salinity, drought and heavy metal stress during seed germination stage in ricebean [*Vigna umbellata* (Thunb.) Ohwi and Ohashi]. *Plant Physiology Reports*, 26(1), 109-115. <https://doi.org/10.1007/s40502-020-00542-4>.
- Baran, M.S. & Kocabağlı, N. (2000). Tane sorgumun süt ineklerinde ruminal fermentasyon, süt verimi ve sütün bileşimi üzerine etkisi. *İstanbul Üniversitesi Veterinerlik Fakültesi Dergisi*, 26, 113-128.
- Batista, V.A.P., Vieira, H.D., Pires, J.I.C., & Correia, L.Z. (2021). Physical-physiological quality and early performance of sorghum plants under different boron doses via seed. *Semina: Ciências Agrárias*, 42(6), 3185-3200.
- Bhamburdekar, S.B. (2002). *Germination studies in pigeon pea (Cajanus cajan)* [Ph.D. Thesis, Shivaji University Kolhapur, India. ] Pp.112
- Bhati, S., Chaudhary, S., & Garg, G. (2021). Effect of soil salinity on growth parameters and antioxidant activity in two genotypes of eggplant (*Solanum melongena* L.). *International Journal of Agricultural and Applied Sciences*, 2(2), 95-102.
- Bibi, A., Zahid, M.I., Sadaqat, H.A., & Fatima, B. (2016). Correlation analysis among forage yield and quality components in sorghum sudangrass hybrids under water stress conditions. *G.J.B.B.*, 5(4), 444-448.
- Carpıcı, E.B., Celik, N., & Bayram, G. (2009). Effects of salt stress on germination of some maize (*Zea mays* L.) cultivars. *African Journal of Biotechnology*, 8(19), 4918-4922.
- Ching, T.M. (1959). Activation of germination in douglas-fir seed by hydrogen peroxide. *Plant Physiol.*, 34, 557-563.
- Datta, J.K., Nag, S., Banerjee, A., & Mondal, N.K. (2009). Impact of salt stress on five varieties of wheat *Triticum aestivum* L. cultivars under laboratory condition. *J. Applied Sci Environ Manage*, 13, 93-97.
- Da Rocha Pinho, L.G., Monnerat, P.H., Pires, A.A., Freitas, M.S.M., & Marciano, C.R. (2015). Diagnosis of boron deficiency in green dwarf coconut palm. *Agric. Sci.*, 6, 164-174.
- dos Santos, L.M., de Farias, S.G.G., e Silva, R.B., Dias, B.A.S., & da Silva, L.S. (2019). Ecophysiology of germination of *Parkia platycephala* Benth. seeds. *Floresta e Ambiente*, 26(1), 1-7. <https://doi.org/10.1590/2179-8087.028215>.
- Ejeta, G. & Knoll, J.E., (2007). *Marker-Assisted Selection in Sorghum In: Genomic-Assisted Crop Improvement: Genomics Applications in Crops*. Varshney RK, Tuberosa Reds. 2, 187-205
- El-Kassaby, Y.A., Moss, I., Kolotelo, D., & Stoehr, M. (2008). Seed Germination: Mathematical Representation and Parameters Extraction. *Forest Science*, 54(2), 220-227.
- El-Kassaby, Y.A. (2000). Representation of douglas-fir and western hemlock families in seedling crops as affected by seed biology and nursery crop management practices. *For. Genet.*, 7,305-315.
- Guo, X., Zhi, W., Feng, Y., Zhou, G., & Zhu, G. (2022). Seed priming improved salt-stressed sorghum growth by enhancing antioxidative defense. *PLoS ONE*, 17(2), e0263036. <https://doi.org/10.1371/journal.pone.0263036>.
- Gupta, U., & Solanki, H. (2013). Impact of boron deficiency on plant growth. *International Journal of Bioassays*, 2(7), 1048-1050.
- Haileselasie, T.H., & Gselasie, B. (2012). The Effect of salinity (NaCl) on germination of selected grass pea (*Lathyrus sativus* L.) landraces of tigray. *Asian Journal of Agricultural Sciences*, 4(2), 96-101.
- Hu, H., & Brown, P.H. (1997). *Absorption of Boron by Plant Roots*. In: Boron in Plants and Soils. Kluwer Acad. Publ., Dordrecht, The Netherlands: 49-58.
- ISTA, 2017. International for Seed Testing Rules. International Seed Testing Association, Zurich, Switzerland.
- Jadhav, S.S. & Bhamburdekar, S.B. (2014). Effect of boron on germination performance in different varieties of sweet sorghum. *International Journal of Advanced Research*, 2(4),1137-1143.
- Kara, B., Akgün, İ., & Altındal, D. (2011). Tritikale genotiplerinde çimlenme ve fide gelişimi üzerine tuzluluğun (NaCl) etkisi. *Selçuk Tarım Bilimleri Dergisi*, 25, 1-9.
- Karadağ, Y., & Özkurt, M. (2014). İkinci ürün olarak yetiştirilebilecek silajlık sorgum (*Sorghum bicolor*



- L. Moench) çeşitlerinde farklı sıra aralıklarının verim ve kalite üzerine etkisi. *Gaziosmanpaşa Üniversitesi Ziraat Fakültesi Dergisi*, 31, 19-24.
- Karle, S.B., Guru, A., Dwivedi, P., & Kumar, K. (2021). Insights into the role of gasotransmitters mediating salt stress responses in plants. *Journal of Plant Growth Regulation*, 0123456789. <https://doi.org/10.1007/s00344-020-10293-z>.
- Kaya, M.D., Ipek, A., & Ozturk, A. (2003). Effects of different soil salinity levels on germination and seedling growth of safflower (*Carthamus tinctorius* L.). *Turk. J. Agric.For.*, 27, 221-227.
- Kebede, H., Subudhi, P.K., Rosenow, D.T., Nguyen, H.T. (2001). Quantitative trait loci influencing drought tolerance in grain sorghum (*Sorghum bicolor* L. moench). *Theor. Appl. Genet.*, 103, 266-276.
- Krishnamurthy, L., Serraj, R., Hash, C.T., Dakheel, A.J., & Reddy, B.S.V. (2007). screening sorghum genotypes for salinity tolerant biomass production. *Euphytica*, 156 (1-2), 15- 24.
- Kumar, D., Al Hassan, M., Naranjo, M.A., Agrawal, V., Boscaiu, M., & Vicente, O. (2017). Effects of salinity and drought on growth, ionic relations, compatible solutes and activation of antioxidant systems in oleander (*Nerium oleander* L.). *Plos One*. 12(9), e0185017. <https://doi.org/10.1371/journal.pone.0185017>. PMID: 28922384.
- Li, W., Yamaguchi, S., Khan, M.A., An, P., Liu, X., & Tran, L.S.P. (2016). Roles of gibberellins and abscisic acid in regulating germination of suaeda salsa dimorphic seeds under salt stress. *Frontiers in Plant Science*, 6, 1-10.
- Maas, E.V., & Hoffman, G.J. (1977). Crop salt tolerance-current assessment. journal of irrigation drainage division. *American Society Civil Engineering*, 103, 115-134.
- Majda, C., Khalid, D., Aziz, A., Rachid, B., Badr, A.S., Lotfi, A., & Mohamed, B. (2019). Nutri-priming as an efficient means to improve the agronomic performance of molybdenum in common bean (*Phaseolus vulgaris* L.). *Science of the Total Environment*, 661, 654-663.
- Marschner, P. (2012). *Mineral Nutrition Of Higher Plants* (3rd ed.). Oxford: Elsevier.
- Moghaddam, M., Farhadi, N., Panjtandoust, M., & Ghanati, F. (2020). Seed germination, antioxidant enzymes activity and proline content in medicinal plant tagetes minuta under salinity stress. *Plant Biosystems*, 154(6), 835-842. <https://doi.org/10.1080/11263504.2019.1701122>.
- Nawaz, F., Ashraf, M.Y., Ahmad, R., Waraich, E.A., & Shabbir, R.N. 2014. Selenium (Se) regulates seedling growth in wheat under drought stress. *Hindawi Publishing Corporation. Advances in Chemistry*, Article ID 143567; 7p.
- Ödemiş, B., Büyüktaş, D., & Çalışkan, M.E. (2019). Effects of saline irrigation water and proline applications on yield, vegetative and physiological characteristics of potato crop (*Solanum tuberosum* L.). *Horticultural Studies* (HortiS), 36, 54-63. <http://doi.org/10.16882/derim.2018.407736>.
- Önal Aşçı, Ö., & Üney, H. (2016). Farklı tuz yoğunluklarının macar fiğinde (*Vicia pannonica* Crantz) çimlenme ve bitki gelişimine etkisi. *Akademik Ziraat Dergisi*, 5, 29-34.
- Pirasteh-Anosheh, H., Emam, Y., Rousta, M.J., & Ashraf, M. (2017). Salicylic acid induced salinity tolerance through manipulation of ion distribution rather than ion accumulation. *Journal of Plant Growth Regulation*, 36, 227-239.
- Pirasteh-Anosheh, H., & Hedayati-Firoozabadi, A. (2022). Sorghum [*Sorghum bicolor* (L.) Moench.] growth, and soil moisture and salt content as affected by irrigation water salinity. *International Journal of Applied and Experimental Biology*, 1(1), 33-37.
- Pirasteh-Anosheh, H., Ranjbar, G., Pakniyat, H., & Emam, Y. (2016). *Physiological Mechanisms of Salt Stress Tolerance in Plants: An Overview*: Pp. 141-160. In: Azooz, M.M., Ahmad, P. (Eds). "Plant-Environment Interaction: Responses and Approaches to Mitigate Stress". Wiley, London.
- Pontieri, P., Di Fiore, R., Troisi, J., Bean, S.R., Roemer, E., Okot, J., Alifano, P., Pignone, D., Giudice, L.D., & Massardo, D.R. (2012). Chemical composition and fatty acid content of white food sorghums grown in different environments. *Maydica*, 56(1), 51
- Quamruzzaman, M., Manik, S.N., Livermore, M., Johnson, P., Zhou, M., & Shabala, S. (2022). Multidimensional screening and evaluation of morpho-physiological indices for salinity stress tolerance in wheat. *Journal of Agronomy and Crop Science*. 208(4), 454-471.
- Rajakumar, R. (2013). A study on effect of salt stress in the seed germination and bio chemical parameters of rice (*Oryza sativa* L.) under in vitro condition. *Asian Plant Sci. Res.*, 3(6), 20-25.
- Reguera, M., Espi, A., Bolanos, L., Bonilha, I., & Redondonieto, M, (2009). Endoreduplication before cell differentiation fails in boron-deficient legume nodules. is boron involved in signaling during cell cycle regulation? *New Phytologist*, 183(1), 8-12. <https://doi.org/10.1111/j.1469-8137.2009.02869.x>.
- Ren, Y., Wang, W., He, J., Zhang, L., Wei, Y., & Yang, M. (2020). Nitric oxide alleviates salt stress in seed germination and early seedling growth of pakchoi (*Brassica chinensis* L.) by enhancing physiological and biochemical parameters. *Ecotoxicology and Environmental Safety*, 187, 109785. <https://doi.org/10.1016/j.ecoenv.2019.109785>.
- Rooney, L.W., & Waniska, R.D, (2000). *Sorghum food and industrial utilization*, pp. 689-729. In: Smith CW, Frederiksen RA eds. Sorghum: Origin, History, Technology, and Production, John Wiley & Sons Inc., New York.

- Shabala, S. (2009). Salinity and programmed cell death: unravelling mechanisms for ion specific signalling. *Journal of Experimental Botany*, 60(3), 709–712. <https://doi.org/10.1093/jxb/erp013>.
- Shahid, M., Ameen, F., Maheshwari, H.S., Ahmed, B., AlNadhari, S., & Khan, M.S. (2021). Colonization of *Vigna radiata* by a halotolerant bacterium *Kosakonia sacchari* improves the ionic balance, stressor metabolites, antioxidant status and yield under NaCl stress. *Applied Soil Ecology*, 158(November 2020), 103809. <https://doi.org/10.1016/j.apsoil.2020.103809>.
- Shakeri, E., Emam, Y., Tabatabaei, S., & Sepaskhah, A. (2017). Evaluation of grain sorghum (*Sorghum bicolor* L.) lines/cultivars under salinity stress using tolerance indices. *Int. J. Plant Prod.*, 11, 101–115.
- Shtaya, M.J., Al-Fares, H., Qubbaj, T., Abu-Qaoud, H., & Shraim, F. (2021). Influence of salt stress on seed germination and agromorphological traits in chickpea (*Cicer arietinum* L.). *Legume Research-An International Journal*, 44(12), 1455-1459.
- Silva-Matos, R.R.S.D., Albano, F.G., Cavalcante, Í.H.L., Pessoa, J.A., Neto Silva, R.L., Oliveira, I.V.D.M., & Carvalho, C.I.F.S. (2017). Desenvolvimento inicial de mudas de melancia cv. crimson sweet em função de doses de boro aplicadas na semente. *Revista de Ciências Agrárias*, 40(4), 30-39. doi: 10.19084/RCA16121.
- Taiz, L., Zeiger, E., Moller, I.M., & Murphy, A. (2015). *Plant Physiology and Development*. Sinauer Associates, Sunderland, Massachusetts, United States.
- Tariq, M., & Mott, C.J. (2006). Effect on boron supply on the uptake of micronutrients by radish (*Raphanus sativus* L.). *J. of Agri. and Bio.Sci.*, 1(2), 1-8.
- Tenikecier, H.S., & Ates, E. (2022). Impact of salinity on germination and seedling growth of four cool-season turfgrass species and cultivars. *Pol. J. Environ. Stud.*, 31(2), 1813-1821.
- Thomson, A.J., & El-Kassaby, Y.A. (1993). Interpretation of seed-germination parameters. *New For.* 7, 123–132.
- Tigabu, E., Andargie, M., & Tesfaye, K. (2012). Response of sorghum (*Sorghum bicolor* (L.) Moench) genotypes to NaCl levels at early growth stages. *Afr. J. Agric. Res.*, 7, 5711–5718.
- Tolay, I. (2021). The impact of different zinc (Zn) levels on growth and nutrient uptake of basil (*Ocimum basilicum* L.) grown under salinity stress. *Plos One*, 16(2), e0246493. <https://doi.org/10.1371/journal.pone.0246493>.
- Ulusik, I., Karakaya, H.C., & Koc, A. (2018). The importance of boron in biological systems. *Journal of Trace Elements in Medicine and Biology*, 45(1), 156-162. <https://doi.org/10.1002/9781119487210.Ch20>.
- Uraguchi, S., & Fujiwara, T. (2011). Significant contribution of boron stored in seeds to initial growth of rice seedlings. *Plant and soil*, 340(1), 435-442. <https://doi.org/10.1007/s11104-010-0614-9>.
- Vadakekara Joseph, M. (2016). Extrusion, physiochemical characterization and nutritional evaluation of sorghum-based high protein, micronutrient for TIFIED. *Blended Foods*. 235 pp.
- Warrington, K. (1923). The effect of boric acid and borax on the broad bean and certain other plants. *Ann. Bot. (Lond.)*, 37, 629-672.
- Xia, F.S., Wang, Y.C., Zhu, H.S., Ma, J.Y., Yang, Y.Y., Tian, R., & Dong, K.H. (2019). Influence of priming with exogenous boron on the seed vigour of alfalfa (*Medicago sativa* L.). *Legume Research-an International Journal*, 42(6), 795-799.
- Yildirim, E., Karlidag, H., & Dursun, A. (2011). Salt tolerance of physalis during germination and seedling growth. *Pakistan Journal of Botany*, 43, 2673-2676.