



Protective Effects of Hesperidin and Salicylic Acid on *Lemna minor* L. Exposed to Evercion Yellow Textile Dye

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ABSTRACT

Hesperidin (HES) is a flavonone glycoside from the flavonoid family that is present in citrus species. It has potent anti-oxidant and anti-cancer properties. In times of stress, the phenolic chemical salicylic acid (SA), also known as a plant hormone, functions as a signal molecule, controlling the plant's reaction and maintaining its survival. For the removal of numerous harmful chemicals, phytoremediation, sometimes referred to as green reclamation, is an efficient, affordable, environmentally benign, and simple procedure. Duckweed (*Lemna minor* L.) is an important bioindicator species in phytoremediation study. Following the application of 75 ppm, 150 ppm, and 300 ppm reactive dye Evercion yellow 1X, the effects of 0.5 mM SA and 0.5 mM hesperidin on duckweed (*L. minor* L.) were examined in this study. The use of 0.5 mM SA against stress boosted the activities of peroxidase (POD), ascorbate peroxidase (APX), glutathione S-transferase (GST), glutathione reductase (GR), superoxide dismutase (SOD), and catalase (CAT). Additionally, total glutathione (GSH), total chlorophyll, and carotenoid content were altered by SA treatment. Similar to the SA application, the application of HES was effective in lowering stress. Lipid peroxidation content measured as malondialdehyde (MDA) content was found to be higher than the control groups. Results suggest that SA plays a positive role in *L. minor* against Evercion yellow 1X.

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Evercion Sarı Tekstil Boyasına Maruz Kalan *Lemna minor* L. Üzerinde Hesperidin ve Salisilik Asit'in Koruyucu Etkileri

ÖZET

Hesperidin (HES), turuncgil türlerinde bulunan flavonoid ailesinden bir flavonon glikozittir. Güçlü anti-oksidan ve anti-kanser özelliklere sahiptir. Stres zamanlarında, bitki hormonu olarak da bilinen fenolik kimyasal salisilik asit (SA) bir sinyal molekülü olarak işlev görür, bitkinin tepkisini kontrol eder ve hayatta kalmasını sağlar. Çok sayıda zararlı kimyasalın ortadan kaldırılması için, bazen yeşil ıslah olarak da adlandırılan fitoremediasyon, verimli, uygun maliyetli, çevreye zarar vermeyen ve basit bir prosedürdür. Su mercimeği (*Lemna minor* L.) fitoremediasyon çalışmalarında önemli bir biyoindekatör türüdür. Bu çalışmada, 75 ppm, 150 ppm ve 300 ppm reaktif boya Evercion yellow 1X uygulamasının ardından, 0.5 mM SA ve 0.5 mM HES'in su mercimeği (*L. minor* L.) üzerindeki etkileri incelenmiştir. Strese karşı 0,5 mM SA kullanımı peroksidaz (POD), askorbat peroksidaz (APX), glutatyon S-transferaz (GST), glutatyon redüktaz (GR), süperoksit dismutaz (SOD) ve katalaz (CAT) aktivitelerini artırmıştır. Ayrıca, toplam gutatyon (GSH), toplam klorofil ve karotenoid içeriği SA uygulaması ile değişmiştir. Malondialdehit (MDA) içeriği olarak ölçülen lipid peroksidasyon düzeyi, kontrol gruplarına göre daha yüksek bulunmuştur.

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INTRODUCTION

The textile industry is one of the world's main sources of pollution because it releases undesirable dye effluents (Yaseen & Scholz, 2019). Reactive dyestuffs are widely used in the textile industry because of their ease of use and stability. Both organic and inorganic pollutants can have a considerable negative impact on plant growth (Purwanti et al. 2018; Titah et al. 2018; Tangahu et al., 2019; Imron et al., 2019). Plants as bioremediation agents hold great promise for the breakdown of various hues as well as organic and inorganic contaminants (Ali, 2010; Singh & Singh, 2017).

A member of the Lemnaceae family, duckweed is a little and uncomplicated floating aquatic plant (Driever et al., 2005; Ceschin et al., 2018). *L. minor* is a useful plant for phytoremediation because it can quickly develop, adapt to a variety of aquatic circumstances, and collect pollutants (Ceschin et al., 2020; Can Terzi et al., 2021). Numerous physiological activities, such as gene transcription, signal transduction, redox signal pathways, and immunological response, require reactive oxygen species (ROS). However, excessive ROS generation results in damaging effects on DNA, proteins, and lipids that ultimately cause cell death (Li et al., 2017). Plants have developed defense mechanisms, both enzymatic and non-enzymatic, to scavenge ROS and lessen their harmful effects (Singh & Satheeshkumar, 2024).

Hesperidin is a flavanone that can be found in citrus and other plants naturally. It has been shown to have a variety of pharmacological actions, including antioxidant, anti-inflammatory, anti-hypercholesterolemic, and anticarcinogenic qualities (Tamilselvam et al., 2013; Janigashvili et al., 2024).

SA is a natural plant growth regulator that has a significant impact on a variety of physiological and metabolic processes that affect a plant's ability to grow and develop. Therefore, SA can be used to counteract elemental toxicities, heavy metal stressors, pathogen virulence, and salt stress (Wani et al., 2017).

The purpose of this study was to determine how SA and hesperidin protected *L. minor* from a reactive dye called Evercion Yellow 1X (EY). Antioxidant enzymes, pigment analysis, and lipid peroxidation tests were performed on samples collected on days 1, 4, and 7.

MATERIAL and METHOD

Hesperidin used in this study was originally synthesized in the chemistry laboratory at Inonu University. The hesperidin compound (Figure 1) was synthesized by organic orange fruits and identified by FT-IR spectroscopy.

Hesperidin synthesis from the orange fruits

Although hesperidin is an insoluble flavonoid in water, its solubility varies depending on pH. First, the dried oranges were ground and powdered. 12.5 g of ground orange powder was dispersed in 125 mL distilled water for 1 hour and mixed with a magnetic stirrer. Since Hesperidin was insoluble in water, the water-soluble species were removed by filtration. CaCO₃ was used to adjust the pH of the environment. 25 g of CaCO₃ was dissolved in 250 ml of distilled water, the Ca(OH)₂ solution obtained by removing the insoluble part was used to adjust the pH of the environment. Ca(OH)₂ solution was added dropwise onto the orange mixture and the pH of the medium was adjusted to 10, and it was mixed for 24 hours to solubilize hesperidin. After solubilization, the mixture was filtered with filter paper, the insoluble part was removed and the pH of the solution was neutralized with HCl. Hesperidin, which dissolves in basic pH but does not dissolve in neutral pH, was obtained by precipitation. Efficiency 85% (Figure 2).

In FTIR spectra (Figure 3), hydroxyl tension peaks at 3400 cm⁻¹, C-H tension peaks of the aromatic ring at 2800 cm⁻¹ circumference, and aromatic C = C tension peaks at 1600 cm⁻¹.

Experimental Design

L. minor mature plants in good condition were obtained from the Turkish seed firm Erciyes in Kayseri. EY, a reactive dyestuff, was selected for this study. Plants were acclimated to Hoagland medium in the greenhouse (temperature: 23 ± 2°C; light/dark cycle: 16/8 h; light intensity: 10,000 lux) for one week before dye treatment (Hoagland & Arnon, 1950). Healthy fronds (30-40 g) from plants were detached and placed in 250 ml glass beakers

with 1/30-diluted Hoagland culture solution with one of the following treatments; Hoagland medium, 75 ppm EY, 150 ppm EY, 300 ppm EY, 0.5 mM SA, 0.5 mM SA + 75 ppm EY, 0.5 mM SA + 150 ppm EY, 0.5 mM SA + 300 ppm EY, 0.5 mM HES, 0.5 mM HES + 75 ppm EY, 0.5 mM HES + 150 ppm EY and 0.5 mM HES + 300 ppm EY. Each experiment was replicated three times. The fronds were picked on days one, four, and seven.

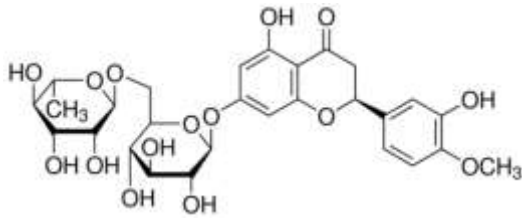


Figure 1. Hesperidin molecule
Şekil 1. Hesperidin molekülü



Figure 2. Hesperidin
Şekil 2. Hesperidin

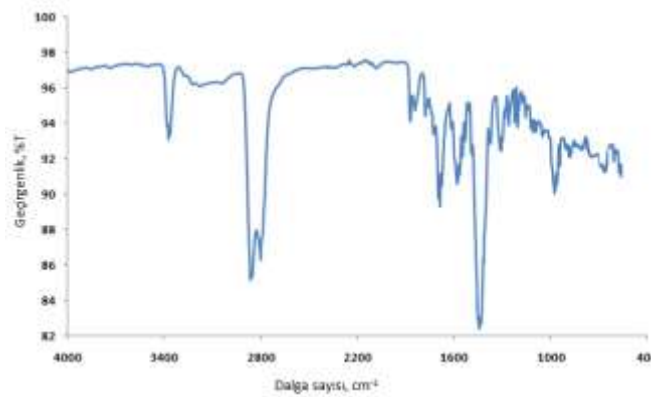


Figure 3. FT-IR spectrum of hesperidin
Şekil 3. Hesperidin FT-IR spektrumu

Enzyme extraction and protein content

According to Huang et al. (2013), enzyme extractions were evaluated. A 5 ml solution of 0.1 M potassium phosphate buffer (pH 7.8) was used to homogenize 0.5 g of *L. minor*. At 4°C and 10000 g, homogenates were centrifuged for 15 minutes. Using bovine serum albumin as the reference protein, the protein content was assessed using a Bradford protein assay (Bradford, 1976).

POD activity

POD activity was calculated in accordance with Peters et al. (1989) and Mac Adam et al. (1992). A solution was prepared by vortexing 3 ml of 0.1 M potassium phosphate buffer (pH 6.0), 0.04 ml of 0.03 M H₂O₂, and 0.05 ml of 0.2 M guaiacol. To 0.9 mL of this solution, 0.1 mL of extract was added. The increase of absorbance at 436 nm was recorded within 1 min. The extinction coefficient of guaiacol is 26.6 mM⁻¹ cm⁻¹.

APX activity

The measurement of APX activity was followed Nakano and Asada's (1981) guidelines. The reaction mixture was prepared by adding 550 µl of phosphate buffer (pH 7.6), 100 µl of 10 mM EDTA, 12 mM H₂O₂, 250 µl of extract, and 100 µl of 0.25 mM AsA. Enzyme activity was determined as the absorbance change obtained in 1 minute at a wavelength of 290 nm. APX activity was calculated with an extinction coefficient of 2.8 mM⁻¹cm⁻¹.

GST activity

The GST activity was determined using the Habig et al. (1974) technique. The reaction mixture included 400 µl of 0.1 M potassium phosphate buffer, 400 µl of reduced glutathione, 100 µl of sample, and 150 µl of CDNB. The absorbance change over 1 minute at a wavelength of 344 nm was used to determine enzyme activity. CDNB has an extinction coefficient of 9.6 mM⁻¹ cm⁻¹.

GR activity

According to Carlberg & Mannervik (1985), an assessment of GR activity was carried out. A solution containing 2

mM EDTA in 0.2 M potassium phosphate buffer (pH 7), 10 mM Tris-HCl (pH 7), 2 mM NADPH, and 20 mM GSSG in 10 ml distilled water was prepared. The reaction mixture was prepared by combining 500 µl of 0.2 M potassium phosphate buffer, 50 µl of 2 mM NADPH, 50 µl of 20 mM GSSG, 250 µl of distilled water, and 150 µl of the sample. The enzyme activity was measured as the change in absorbance per minute at 340 nm and calculated using the NADPH extinction coefficient of $6.2 \text{ mM}^{-1} \text{ cm}^{-1}$.

GSH content

The amount of GSH was calculated in accordance with Akerboom & Sies (1981). 2.378 mg of DTNB and 0.248 mg of NADPH were prepared in 1 ml of 125 mM sodium diphosphate buffer (pH 7.5) containing 6.3 mM EDTA. 700 µl of NADPH and 100 µl of DTNB were incubated in a water bath at 30 °C, then 100 µL of distilled water, 5 ml of GR, and 100 µl of the extract were added. The reaction was calculated at 412 nm.

SOD activity

The McCord Fridovich (1969) approach was used to assess SOD activity. A solution A was prepared by mixing 100 mL of a 50 mM K_2HPO_4 buffer at pH 7.8 with a solution containing 24.8 mg of cytochrome c and 10 mL of distilled water with 0.76 mg of xanthine. Solution B was prepared separately by dissolving 0.2 U/ml of xanthine oxidase. A reaction mixture was prepared by combining 900 µl of solution A, 50 µl of solution B, and 100 µl of the sample. At 550 nm, enzyme activity was determined.

CAT activity

The Luck (1965) technique was used to evaluate CAT activity. A 1/15 M pH 7 potassium phosphate buffer has been prepared, and 160 µl of H_2O_2 was added to 100 ml of this buffer. Then 200 µl of sample was added to 800 µl of phosphate buffer to which H_2O_2 was added, and absorbance changes over 1 minute were determined by reading at 240 nm. The molar extinction coefficient for H_2O_2 is $0.0396 \text{ cm}^2 \mu\text{mol}^{-1}$.

Pigments analysis

De Kok and Graham's (1980) approach was utilized to determine pigment content. To begin, a 1 g leaf sample was homogenized in 50 ml of acetone and stored at +4°C for 24 hours. After adding 1/5 water to the homogenate, it was homogenized again in a shaking incubator for 15 minutes and kept at +4 °C for 24 hours. At the end of the period, the samples were centrifuged at 5000 g for 10 minutes. The absorbance values of the centrifuged samples were read in a spectrophotometer at 662, 645, 470 nm according to Lichtenthaler and Welburn (1983).

Lipid peroxidation assay

Heath and Packer (1968) used the thiobarbituric acid reactive chemicals approach to measure MDA levels. A 0.5 g leaf sample was homogenized with 5 mL of 0.1% trichloroacetic acid (TCA), and the homogenate was centrifuged at 10,000 g for 10 minutes. From the centrifuged sample, 2 mL of the supernatant was taken and 2 mL of 0.5% thiobarbituric acid (TBA) was added. The mixture was incubated in a water bath at 95°C for 30 minutes. The absorbance was measured using both 532 and 600 nm wavelengths. MDA content was calculated using an extinction coefficient of $155 \text{ mM}^{-1} \text{ cm}^{-1}$.

Statistical Analysis

The results were reported as the mean with standard error (SE). The entire data set from all studies was statistically evaluated using SPSS version 25.0. In the study, the conformity of the data to normal distribution was checked with the Shapiro-Wilk Test, and the homogeneity of the variances was checked with the LEVENE test. Since the data provided normal distribution, the ANOVA test, one of the parametric test methods, was used in the analysis of multi-group variables and the Duncan multiple comparison test was used for pairwise comparisons between groups (Duncan, 1955).

RESULTS and DISCUSSION

Antioxidant Enzyme Activities

The synthetic dyes used in the textile industry pollute a significant amount of water. Textile dyes do not adhere tightly to the fabric and are released as effluent into the aquatic environment (Rania et al., 2022).

In nature, plants are continually exposed to abiotic and biotic stresses. Furthermore, plants have both enzymatic and non-enzymatic antioxidant defense systems that help to minimize ROS production and cellular damage.

Lemnaceae are among the world's tiniest and quickest growing angiosperms, producing massive amounts of biomass. Lemna species have a high economic value and diverse applications in biotechnology and ecology (Alp et

al., 2023). Their morphological and physiological characteristics enable them to perform valuable bioassays in restricted environments in a short amount of time, making them ideal laboratory organisms. Lemna species are used as bioindicators for ecotoxicological studies, both in vivo and in vitro (Basiglioni et al., 2018; Beker Akbulut et al., 2020; Beker Akbulut & Özhan Turhan, 2021). HES is one of the most common flavonoids discovered in citrus species. It is an active pharmacological compound having antihypertensive, anti-inflammatory, and anticancer properties (Iranshahi et al., 2015; Tabeshpour et al., 2020; Arikan et al., 2022; Ji et al., 2024).

POD is a common enzyme found in animals, plants, microbes, and cultured cells. It catalyzes the oxidation of phenols and amines with hydrogen peroxide, removing their toxicity. POD activity was shown to be higher in the EY-applied groups than in the control group. It rose with time in those treated with 0.5 mM SA and dye. The 0.5 mM HES and dye groups demonstrated an increase in POD activity over time. In the 0.5 mM SA +300 ppm dye groups, the maximal POD activity was 15.42 U/mg protein. The groups treated with 0.5 mM HES +150 ppm dye showed the highest POD activity (16.33 U/mg protein). The changes in POD activity were found to be statistically significant, $F(11,24)= 2.85$, $p=.015$. (Figure 4).

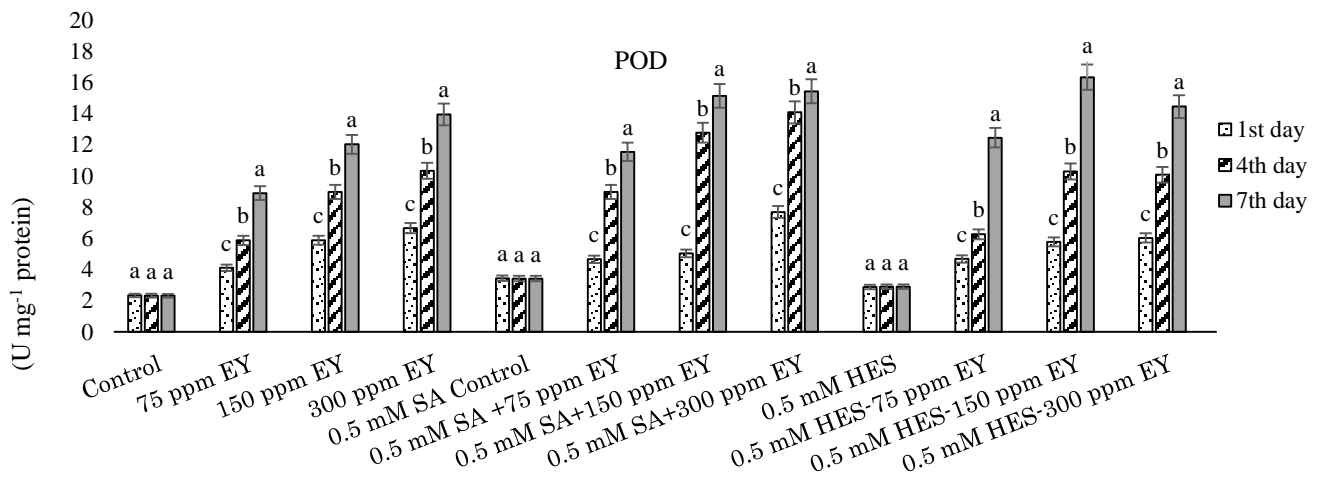


Figure 4. POD activity in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan's test, $F(11,24) = 2.85$, $p=.015$

Şekil 4. EY, SA ve HES uygulanan *L. minor*'da POD aktivitesi. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır, $F(11,24)= 2.85$, $p=.015$

Ascorbate peroxidase enzymes serve a critical function in regulating ROS in cells, particularly H_2O_2 . Following 75, 150, and 300 ppm EY treatment, APX activity increased with the number of days. The 0.5 mM SA and dye administered groups increased as the number of days increased. On the seventh day, the maximum APX activity (15.26 U/mg protein) was identified in the groups treated with 0.5 mM HES (Figure 5).

GSTs are multifunctional proteins that are involved in a variety of intracellular events such as stress metabolism, primary and secondary metabolism, herbicide detoxification, plant defense against heavy metals, ozone destruction, and xenobiotics. GST activity increased with increasing dye concentrations and days. The changes in GST activity were found to be statistically significant, $F(11,24)= 3.57$, $p<.004$. Only in the dye-applied groups did the maximum GST activity result from 300 ppm EY application. GST activation was found to be greater in the 0.5 mM SA-treated groups than in the dye-only groups. GST activity was shown to be decreased in the 0.5 mM HES treated groups compared to the 0.5 mM SA applied groups (Figure 6).

GR reduces glutathione by using NADPH as a substrate. On the fourth and seventh days, dye-only groups had lower GR activity than the control group. (GR activity decreased on the fourth and seventh days in the groups treated with 0.5 mM SA). The lowest GR activity in the 0.5 mM HES treated groups was seen on the seventh day in the 300 ppm EY applied groups. This is due to the fact that there are notable variations between the administered dose groups and the control, $F(11,24)= 1.36$, $p<.254$ (Figure 7).

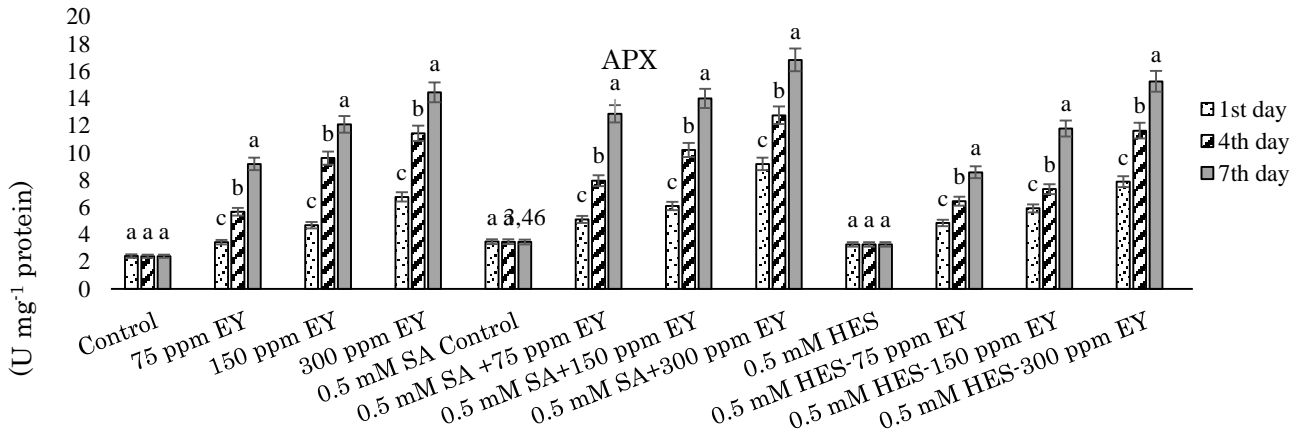


Figure 5. APX activity in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan's test, $F(11,24)= 3.88, p=0.003$

Şekil 5. EY, SA ve HES uygulanan *L. minor*'da APX aktivitesi. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır, $F(11,24)= 3.88, p=0.003$

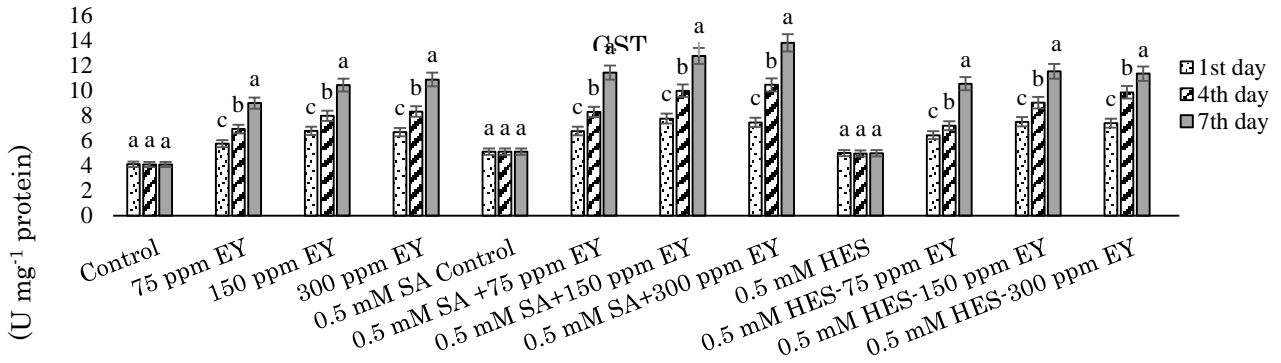


Figure 6. GST activity in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan's test, $F(11,24)= 3.57, p<0.004$

Şekil 6. EY, SA ve HES uygulanan *L. minor*'da GST aktivitesi. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır, $F(11,24)= 3.57, p<0.004$

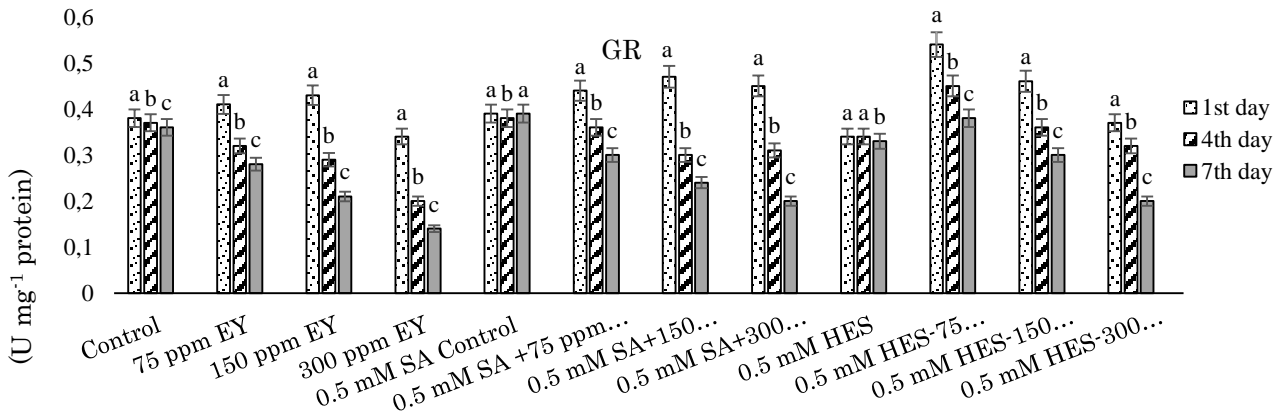


Figure 7. GR activity in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan's test, $F(11,24)= 1.36, p<0.254$

Şekil 7. EY, SA ve HES uygulanan *L. minor*'da GR aktivitesi. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır, $F(11,24)= 1.36, p<0.254$

GSH is a key molecule in sulfur metabolism and plant response to oxidative stress. GSH levels increased in response to increasing dye concentrations and days. The dye-applied groups had the highest GSH concentration. The 0.5 mM SA treated groups had higher GSH content than the dye-only groups. On the seventh day, the groups that applied 0.5 mM HES-300 ppm EY had the highest GSH content (8.36 U/mg protein). This is because significant differences exist between the applied dose groups and the control, $F(211,24)= 6.36, p<.000$ (Figure 8).

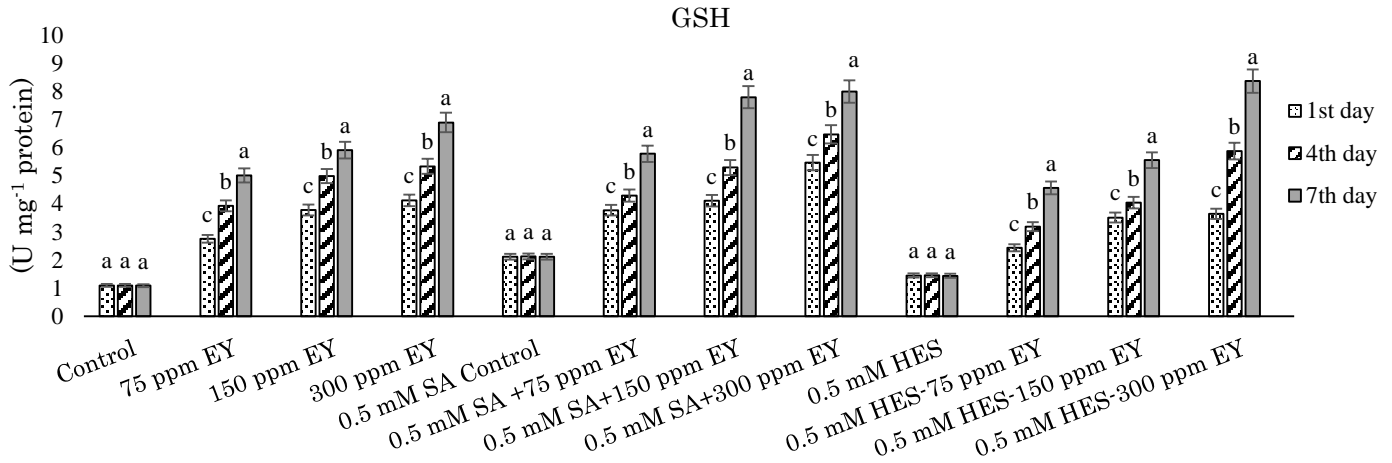


Figure 8. GSH content in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other, $F(11,24)= 6.36, p<.000$

Şekil 8. EY, SA ve HES uygulanan *L. minor*'da GSH içeriği. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır, $F(11,24)= 6.36, p<.000$.

SOD defends plants against abiotic stress-induced ROS. SOD activity increased in dye-treated groups compared to the control. The changes in SOD activity was found to be statistically significant, $F(11,24)= 3.10, p<.010$. The group that applied 300 ppm EY on the seventh day showed the highest SOD activity. Increased concentrations in the SA and HES applied groups resulted in an increase in SOD activity (Figure 9).

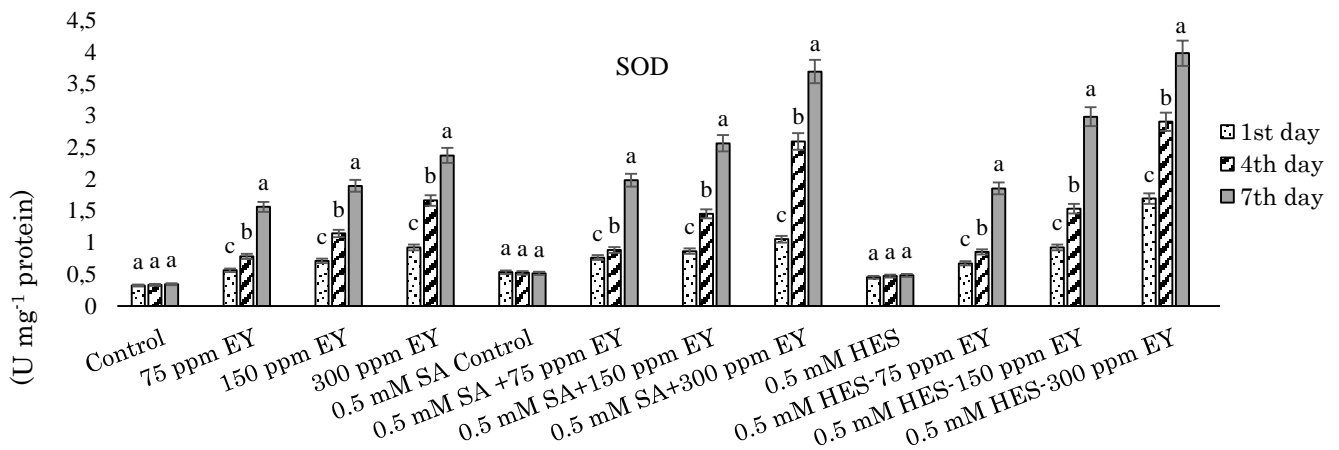


Figure 9. SOD activity in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan's test, $F(11,24)= 3.10, p<.010$

Şekil 9. EY, SA ve HES uygulanan *L. minor*'da SOD aktivitesi. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır $F(11,24)= 3.10, p<.010$

CAT is a tetrameric heme-containing enzyme, removes H_2O_2 from plants under stress to prevent oxidative damage. CAT activity increased on the fourth day solely in the dye-applied groups, but declined on the seventh day. The

groups that applied 300 ppm EY on the seventh day showed the highest CAT activity. On the fourth day, the groups who received both SA and 300 ppm dye showed the highest CAT activity. CAT activity reduced on the seventh day in HES-applied groups. This is because significant differences exist between the applied dose groups and the control, $F(11,24)= 1.94, p<.084$ (Figure 10).

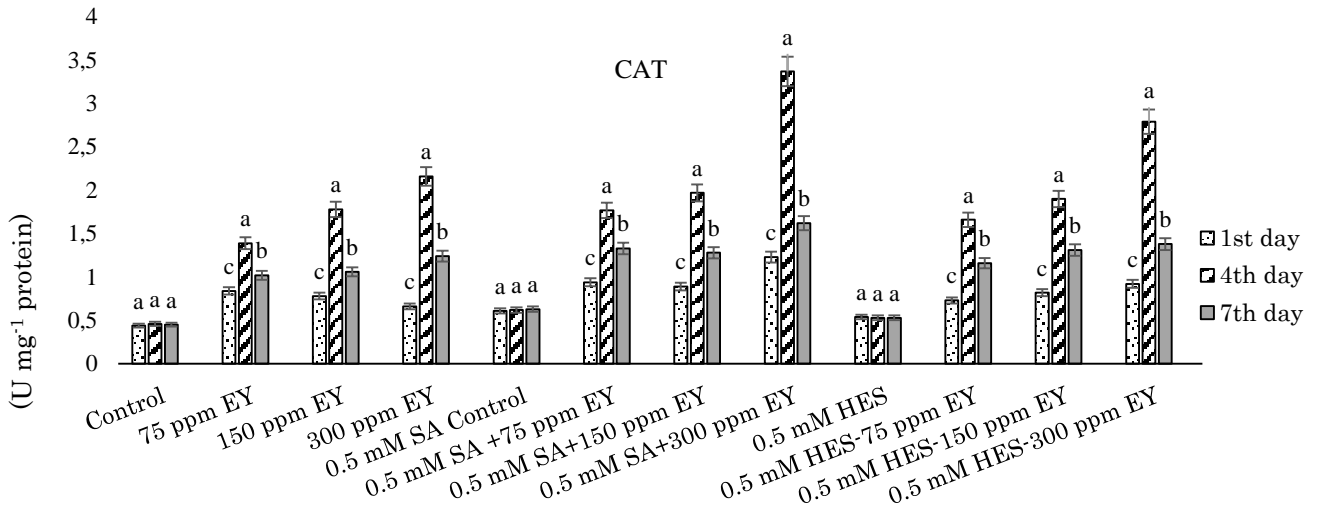


Figure 10. CAT activity in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan’s test, $F(11,24)= 1.94, p<.084$

Şekil 10. EY, SA ve HES uygulanan *L. minor*’da CAT aktivitesi. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır, $F(11,24)= 1.94, p<.084$

Photosynthetic Pigment Analysis

Stress conditions negatively affect photosynthetic activity in plants. In comparison to the control group, the overall chlorophyll concentration in the 75 ppm, 150 ppm, and 300 ppm dye treated groups decreased as time passed. The total chlorophyll content increased on the first day compared to the control, but then decreased on consecutive days in the groups treated with 0.5 mM SA and 75, 150, and 300 ppm dyes. On day 7, total chlorophyll content increased compared to day 4 in groups treated with 0.5 mM HES and 75, 150, or 300 ppm dye. This is due to the fact that there are notable variations between the administered dose groups and the control, $F(11,24)= 11.61, p<.000$ (Figure 11).

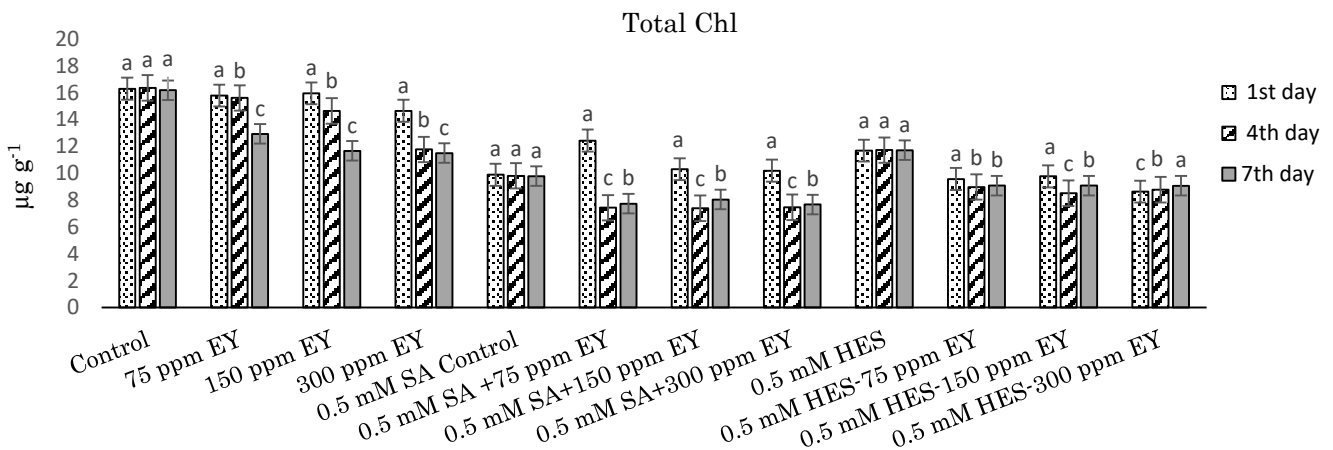


Figure 11. Changes in total chlorophyll content in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan’s test, $F(11,24)= 11.61, p<.000$

Şekil 11. EY, SA ve HES uygulanan *L. minor*’da total klorofil içeriğindeki değişiklikler. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır, $F(11,24)= 11.61, p<.000$

The dye-treated groups' carotenoid concentration reduced by 75 ppm, 150 ppm, and 300 ppm compared to the control. On the first day, groups receiving 0.5 mM SA and 75 ppm dye had the greatest carotenoid levels (4.07 µg g⁻¹). On the seventh day, those treated with 0.5 mM SA and 75 ppm dye had the lowest carotenoid levels (1.78 µg/g). The carotenoid content of the HES treated groups reduced in comparison to the control, with the exception of the HES and 75 ppm dye applied groups on the first day. The changes in carotenoid content were found to be statistically significant, $F(11,24)= 2.24, p<.048$ (Figure 12).

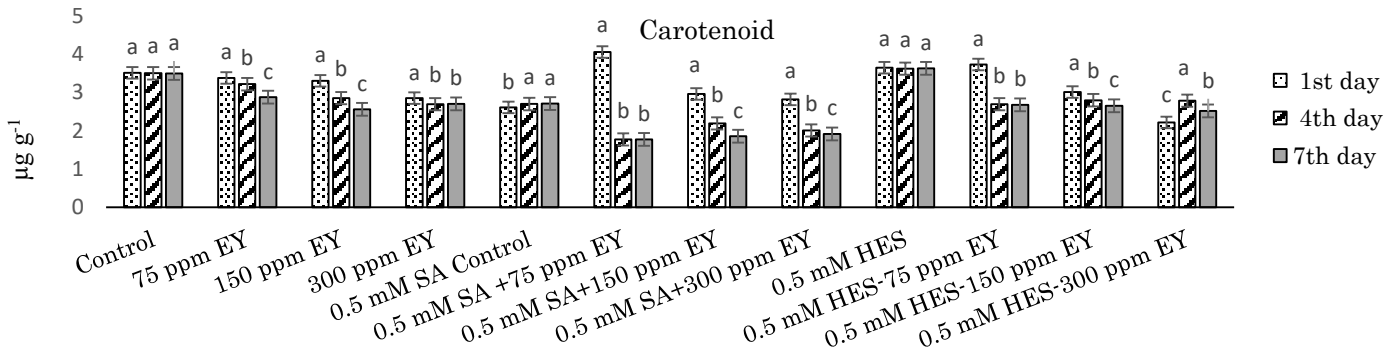


Figure 12. Changes in Carotenoid content in *L. minor* treated with EY, SA and HES. Vertical columns represent the standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan's test, $F(11,24)= 2.24, p<.048$

Şekil 12. EY, SA ve HES uygulanan *L. minor*'da karotenoid miktarındaki değişiklikler. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır, $F(11,24)= 2.24, p<.048$

Lipid Peroxidation Analysis

Increased Lipid peroxidation content in membranes weakens their barrier function and increases permeability to organic substances and ions. This causes sulfhydryl group destruction and enzyme inactivation. On the fourth day, groups that received 300 ppm EY had the highest MDA activity. On the seventh day, MDA levels in the dye-applied groups decreased. MDA concentrations peaked on day four in those treated with 0.5 mM SA and 300 ppm EY. MDA levels in the groups treated with 0.5 mM HES and dye increased on day four but decreased on day seven. This is due to the fact that there are notable variations between the administered dose groups and the control, $F(11,24)= 2.60, p<.025$ (Figure 13).

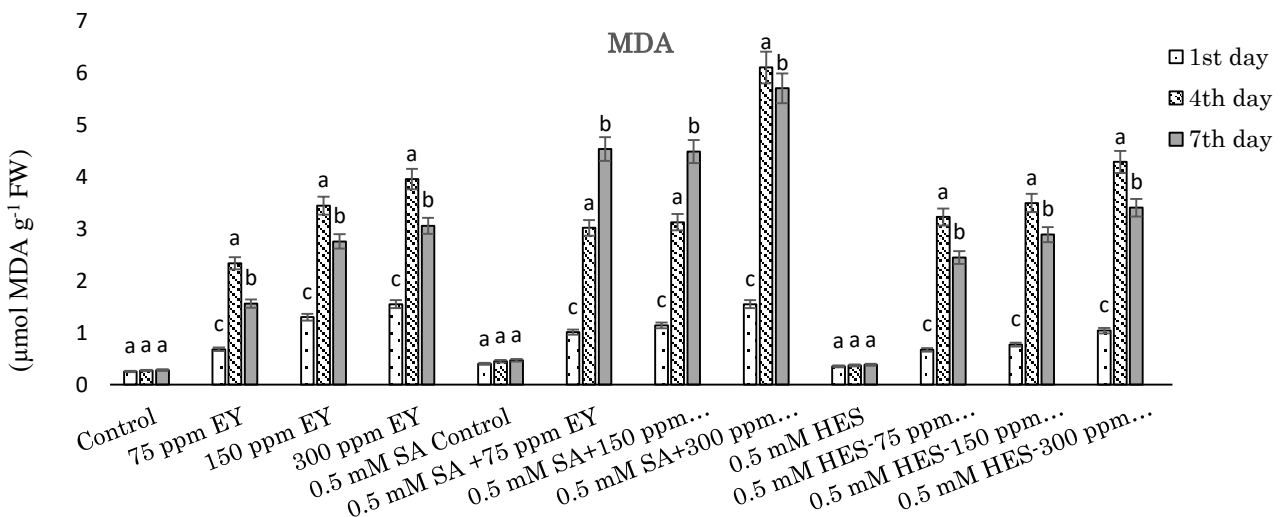


Figure 13. Changes in MDA content in *L. minor* treated with EY, SA and HES. Vertical columns represent standard error of the average of three replications. Data followed by different letters are significantly different from each other according to Duncan's test $F(11,24)= 2.60, p<.025$

Şekil 13. EY, SA ve HES uygulanan *L. minor*'da MDA miktarındaki değişiklikler. Dikey sütunlar üç tekrarın ortalamasının standart hatasını temsil eder. Farklı harflerle takip edilen veriler Duncan testine göre birbirinden anlamlı derecede farklıdır $F(11,24)= 2.60, p<.025$

CONCLUSION

In conclusion, when employed in agriculture, water containing reactive dyes can end up in human bodies. Reactive dyes provide a significant risk to human health and may be a primary contributor to cancerous disorders. A very promising species for natural-environment technologies is *L. minor*. Oxidative stress was brought on by Everzol yellow dye (EY) in *L. minor*. Antioxidant activity and pigmentation were significantly decreased. POD, APX, GST, GSH, and SOD were all elevated upon application of SA and hesperidin. Day 7 saw a decline in GR and CAT activity in the hesperidin and SA groups. Day 4 saw an increase in MDA content. The results of this investigation indicate that the combination of hesperidin and exogenous SA helped *L. minor* recover from dye-induced stress.

Contribution Rate Statement Summary of Researchers

The authors declare that they have contributed equally to the article.

Conflict of Interests and Ethical Statement

The authors of the articles declare that they have no conflict of interest.

REFERENCES

- Akerboom, T.P.M., & Sies, H. (1981). Assay of glutathione, glutathione disulfide and glutathione mixed disulfides in biological samples. *Method Enzymol*, 77, 373-382. [https://doi.org/10.1016/S0076-6879\(81\)77050-2](https://doi.org/10.1016/S0076-6879(81)77050-2).
- Alkimin, G. D., Daniel, D., Dionísio, R., Soares, A. M. V. M., Barata, C., & Nunes, B. (2019). Effects of diclofenac and salicylic acid exposure on *Lemna minor*: Is time a factor? *Environmental Research*, 177, 1-15. <https://doi.org/10.1016/j.envres.2019.108609>.
- Alp, F. N., Arikan, B., Ozfidan-Konakci, C., Ekim, R., Yildiztugay, E., & Turan, M. (2023). Rare earth element scandium mitigates the chromium toxicity in *Lemna minor* by regulating photosynthetic performance, hormonal balance and antioxidant machinery. *Environmental Pollution*, 316, 1-13. <https://doi.org/10.1016/j.envpol.2022.120636>.
- Al-Snai, A. E., (2019). *Lemna minor*: Traditional uses, chemical constituents and pharmacological effects-A review. *IOSR Journal of Pharmacy*, 9(8), 6-11. (e)-ISSN: 2250-3013, (p)-ISSN: 2319-4219.
- Arikan, B., Ozfidan-Konakci, C., Yildiztugay, E., Zengin, G., Alp, F. N., & Elbasan, F. (2022). Exogenous hesperidin and chlorogenic acid alleviate oxidative damage induced by arsenic toxicity in *Zea mays* through regulating the water status, antioxidant capacity, redox balance and fatty acid composition. *Environmental Pollution*, 292, 1-15. <https://doi.org/10.1016/j.envpol.2021.118389>.
- Ashraf, S., Ali, Q., Zahir, Z. A., Ashraf, S., & Asghar, H. N. (2019). Phytoremediation: Environmentally sustainable way for reclamation of heavy metal polluted soils. *Ecotoxicology and Environmental Safety*, 174, 714-727. <https://doi.org/10.1016/j.ecoenv.2019.02.068>.
- Aziz, A., Kapoor, D., 2018. Salicylic Acid: It's physiological role and interactions. *Research Journal of Pharmacy and Technology*, 11(7), 3171-3177. <https://doi.org/10.5958/0974-360X.2018.00583.8>.
- Bansal, N., & Kanwar, S. S. (2013). Peroxidase (s) in environment protection. *The Scientific World Journal*. 1-9. <https://doi.org/10.1155/2013/714639>.
- Beker Akbulut, G., Turhan, D. Ö., & Yiğit, E. (2020). Alleviation of Everzol Red LFB toxicity in duckweed (*Lemna minor* L.) by exogenous salicylic Acid. *KSU J. Agric Nat*, 23(4), 876-884. <https://doi.org/10.18016/ksutarimdogavi.683962>.
- Beker Akbulut, G., Özhan Turhan, D. (2021). Role of salicylic acid in resistance to everzol navy ED in *Lemna minor* L.(Duckweed). *International Journal of Pure and Applied Sciences* 7(1), 185-195. <https://doi.org/10.29132/ijpas.894056>.
- Basiglini, E., Pintore, M., & Forni, C. (2018). Effects of treated industrial wastewaters and temperatures on growth and enzymatic activities of duckweed (*Lemna minor* L.). *Ecotoxicology and Environmental Safety*, 153, 54-59. <https://doi.org/10.1016/j.ecoenv.2018.01.053>.
- Bozbuga, R. (2020). Expressions of pathogenesis related 1 (PR1) gene in *Solanum lycopersicum* and Influence of salicylic acid exposures on host-Meloidogyne incognita interactions. In Doklady. *Biochemistry and Biophysics*. Pleiades Publishing. 494(1), 266-269. <https://doi.org/10.1134/S1607672920050038>.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1-2), 248-254. [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3).
- Buttar, Z. A., Wu, S. N., Arnao, M. B., Wang, C., Ullah, I., & Wang, C. (2020). Melatonin suppressed the heat stress-induced damage in wheat seedlings by modulating the antioxidant machinery. *Plants*, 9(7), 809. <https://doi.org/10.3390/plants9070809>.

- Can-Terzi, B., Goren, A. Y., Okten, H. E., & Sofuoglu, S. C. (2021). Biosorption of methylene blue from water by live *Lemna minor*. *Environmental Technology & Innovation*, *22*, 1-12. <https://doi.org/10.1016/j.eti.2021.101432>.
- Carlberg, I. & Mannervik, B. (1985). Glutathione reductase. *Methods in Enzymology*, *113*, 484-490. [https://doi.org/10.1016/S0076-6879\(85\)13062-4](https://doi.org/10.1016/S0076-6879(85)13062-4).
- Ceschin, S., Crescenzi, M., & Iannelli, M. A. (2020). Phytoremediation potential of the duckweeds *Lemna minuta* and *Lemna minor* to remove nutrients from treated waters. *Environmental Science and Pollution Research*, *27*(13), 15806-15814. <https://doi.org/10.1007/s11356-020-08045-3>.
- Chavoushi, M., Najafi, F., Salimi, A., & Angaji, S. A. (2020). Effect of salicylic acid and sodium nitroprusside on growth parameters, photosynthetic pigments and secondary metabolites of safflower under drought stress. *Scientia Horticulturae*, *259*, 1-6. <https://doi.org/10.1016/j.scienta.2019.108823>.
- Cicerali, N. I. (2004). Effect of stress on antioxidant defense systems of sensitive and resistant cultivars of lentil (*Lens culinaris* M.) (Thesis number 153433). Master's Thesis, ODTÜ, Ankara. Council of High Education Thesis Center.
- Cohen, S. P. & Leach, J. E. (2019). Abiotic and biotic stresses induce a core transcriptome response in rice. *Scientific Reports*, *9*(1), 1-11. <https://doi.org/10.1038/s41598-019-42731-8>.
- De Kok, L. & Graham, M. (1989). Levels of pigments, soluble proteins, amino acids and sulphydryl compounds in foliar tissue of *Arabidopsis thaliana* during dark-induced and natural senescence. *Plant Physiology and Biochemistry* (Paris), *27*(2), 203-209. ISSN/ISBN: 0981-9428.
- Demirci, T., Asci, Ö. A., & Baydar, N. G. (2021). Influence of salicylic acid and L-phenylalanine on the accumulation of anthraquinone and phenolic compounds in adventitious root cultures of madder (*Rubia tinctorum* L.). *Plant Cell, Tissue and Organ Culture*, *144*(2), 313-324. <https://doi.org/10.1007/s11240-020-01952-w>.
- Di, X., Gomila, J. & Takken, F. L. (2017). Involvement of salicylic acid, ethylene and jasmonic acid signalling pathways in the susceptibility of tomato to *Fusarium oxysporum*. *Molecular Plant Pathology*, *18*(7), 1024-1035. <https://doi.org/10.1111/mpp.12559>.
- Dorina, S., Judith, S., Björn, W., Julia, S., Andrea, S., Muffler, K., & Roland, U. (2020). A new strategy for a combined isolation of EPS and pigments from cyanobacteria. *Journal of Applied Phycology*, *32*, 1-12. <https://doi.org/10.1007/s10811-020-02063-x>.
- Duncan, D. B. (1955). Multiple range and multiple F tests. *Biometrics*, *11*(1), 1-42. <http://dx.doi.org/10.2307/3001478>.
- Feng, J., Zhang, M., Yang, K. N., & Zheng, C. X. (2020). Salicylic acid-primed defence response in octoploid strawberry 'Benihoppe' leaves induces resistance against *Podosphaera aphanis* through enhanced accumulation of proanthocyanidins and upregulation of pathogenesis-related genes. *BMC Plant Biology*, *20*, 1-18. <https://doi.org/10.21203/rs.2.16580/v1>.
- Gong, H., Jiao, Y., Hu, W. W., & Pua, E. C. (2005). Expression of glutathione-S-transferase and its role in plant growth and development in vivo and shoot morphogenesis in vitro. *Plant Molecular Biology*, *57*(1), 53-66. <https://doi.org/10.1007/s11103-004-4516-1>.
- Habig, W., Pabst, M., & Jakoby, W. (1974). The first enzymatic step in mercapturic acid formation. Glutathione-S-transferase. *J. Biol. Chem.*, *249*:7130-7139. [https://doi.org/10.1016/S0021-9258\(19\)42083-8](https://doi.org/10.1016/S0021-9258(19)42083-8).
- Hasanuzzaman, M., Nahar, K., Anee, T. I., & Fujita, M. (2017). Glutathione in plants: biosynthesis and physiological role in environmental stress tolerance. *Physiology and Molecular Biology of Plants*, *23*(2), 249-268. <https://doi.org/10.1007/s12298-017-0422-2>.
- Havaux, M. (2014). Carotenoid oxidation products as stress signals in plants. *The Plant Journal*, *79*(4), 597-606. <https://doi.org/10.1111/tpj.12386>.
- Heath, R. L. & Packer, L. (1968). Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. *Archives of Biochemistry and Biophysics*, *125*(1), 189-198. [https://doi.org/10.1016/0003-9861\(68\)90654-1](https://doi.org/10.1016/0003-9861(68)90654-1).
- Hernández-Ruiz, J. & Arnao, M. B. (2018). Relationship of melatonin and salicylic acid in biotic/abiotic plant stress responses. *Agronomy*, *8*(4):33. <https://doi.org/10.3390/agronomy8040033>.
- Hoagland, D. R., Arnon, D. I. (1950). The water culture method for growing plants without soil. UC College of Agriculture, Ag. Exp. Station, Berkeley, CA. *Circular*. *347*, 1-32. https://www.nutricaodeplantas.agr.br/site/downloads/hoagland_arnon.pdf
- Hossen, M. S., Karim, M. F., Fujita, M., Bhuyan, M. B., Nahar, K., Masud, A. A. C., Mahmud J. A., & Hasanuzzaman, M. (2022). Comparative physiology of indica and japonica rice under salinity and drought stress: An intrinsic study on osmotic adjustment, oxidative stress, antioxidant defense and methylglyoxal detoxification. *Stresses*, *2*(2), 156-178. <https://doi.org/10.3390/stresses2020012>.
- Huang, L., Lu, Y., Gao, X., Du, G., Ma, X., Liu, M., & Chen, Y. (2013). Ammonium-induced oxidative stress on plant growth and antioxidative response of duckweed (*Lemna minor* L.). *Ecological Engineering*, *58*, 355-362. <https://doi.org/10.1016/j.ecoleng.2013.06.031>.

- Iatrou, E. I., Kora, E., & Stasinakis, A. S. (2018). Investigation of biomass production, crude protein and starch content in laboratory wastewater treatment systems planted with *Lemna minor* and *Lemna gibba*. *Environmental Technology*, 40(20), 2649-2656. <https://doi.org/10.1080/09593330.2018.1448002>.
- Janigashvili, G., Chkhikvishvili, I., Ratiani, L., Maminaishvili, T., Chkhikvishvili, D., & Sanikidze, T. (2024). Effects and medical application of plant-origin polyphenols: A narrative review. *Bioactive Compounds in Health and Disease-Online ISSN: 2574-0334; Print ISSN: 2769-2426*, 7(8), 375-385. <https://doi.org/10.31989/bchd.v7i8.1414>.
- Ji, Z., Deng, W., Chen, D., Liu, Z., Shen, Y., Dai, J., Zhou H., Zhang, M., Xu, H., & Dai, B. (2024). Recent understanding of the mechanisms of the biological activities of hesperidin and hesperidin and their therapeutic effects on diseases. *Heliyon*. 10(5), <https://doi.org/10.1016/j.heliyon.2024.e26862>.
- Li, Z., Xu, X., Leng, X., He, M., Wang, J., Cheng, S., & Wu, H. (2017). Roles of reactive oxygen species in cell signaling pathways and immune responses to viral infections. *Archives of Virology*, 162(3), 603-610. <https://doi.org/10.1007/s00705-016-3130-2>.
- Li, C., Ji, J., Wang, G., Li, Z., Wang, Y., & Fan, Y. (2020). Over-expression of LcPDS, LcZDS, and LcCRTISO, genes from wolfberry for carotenoid biosynthesis, enhanced carotenoid accumulation, and salt tolerance in tobacco. *Frontiers in Plant Science*, 11:119. <https://doi.org/10.3389/fpls.2020.00119>.
- Lichtenthaler, H. K. & Wellburn, A. R. (1983). Determinations of total carotenoids and chlorophylls a and b of leaf extracts in different solvents: Portland Press Ltd. <https://doi.org/10.1042/bst0110591>.
- Lo, W. J., Chiou, Y. C., Hsu, Y. T., Lam, W. S., Chang, M. Y., Jao, S. C., & Li, W. S. (2007). Enzymatic and nonenzymatic synthesis of glutathione conjugates: application to the understanding of a parasite's defense system and alternative to the discovery of potent glutathione s-transferase inhibitors, *Bioconjugate Chem.*, 18, 109-120. <https://doi.org/10.1021/bc0601727>.
- Luck H. (1965) Catalase. Methods of enzymatic analysis, 885-894. <http://dx.doi.org/10.1016/B978-0-12-395630-9.50158-4>.
- MacAdam, J. W., Nelson, C. J., & Sharp, R. E. (1992). Peroxidase activity in the leaf elongation zone of tall fescue: I. Spatial distribution of ionically bound peroxidase activity in genotypes differing in length of the elongation zone. *Plant Physiology*, 99(3), 872-878. <http://dx.doi.org/10.1104/pp.99.3.872>.
- McCord J.M. & Fridovich I. (1969). Superoxide dismutase: An enzymic function for erythrocyte superoxide (hemocytin), *J. Biol. Chem*, 244(22), 6049-6055. [https://doi.org/10.1016/S0021-9258\(18\)63504-5](https://doi.org/10.1016/S0021-9258(18)63504-5).
- Nakano, Y. & Asada, K. (1981). Hydrogen peroxide is scavenged by ascorbate-specific peroxidase in spinach chloroplasts. *Plant and Cell Physiology*, 22(5), 867-880. <https://doi.org/10.1093/oxfordjournals.pcp.a076232>.
- Parlak, K. U. & Yilmaz, D. D. (2012). Response of antioxidant defences to Zn stress in three duckweed species. *Ecotoxicology and Environmental Safety*, 85, 52-58. <https://doi.org/10.1016/j.ecoenv.2012.08.023>.
- Peters, J. L., Castillo, F. J., & Heath, R. L. (1989). Alteration of extracellular enzymes in pinto bean leaves upon exposure to air pollutants, ozone and sulfur dioxide. *Plant Physiology*, 89(1), 159-164. <https://doi.org/10.1104/pp.89.1.159>.
- Sackey, L. N., Kočí, V., & van Gestel, C. A. (2020). Ecotoxicological effects on *Lemna minor* and *Daphnia magna* of leachates from differently aged landfills of Ghana. *Science of the Total Environment*, 698, 1-7. <https://doi.org/10.1016/j.scitotenv.2019.134295>.
- Sarker, U. & Oba, S. (2020). The response of salinity stress-induced *A. tricolor* to growth, anatomy, physiology, non-enzymatic and enzymatic antioxidants. *Frontiers in Plant Science*, 11, 1-14. <https://doi.org/10.3389/fpls.2020.559876>.
- Sharma, G., & Mathur, V. (2020). Modulation of insect-induced oxidative stress responses by microbial fertilizers in *Brassica juncea*. *FEMS Microbiology Ecology*, 96(4), 1-10. <https://doi.org/10.1093/femsec/fiaa040>.
- Sies, H., Belousov, V. V., Chandel, N. S., Davies, M. J., Jones, D. P., Mann, G. E., Murphy M. P., Yamamoto, M. & Winterbourn, C. (2022). Defining roles of specific reactive oxygen species (ROS) in cell biology and physiology. *Nature Reviews Molecular Cell Biology*, 23(7), 499-515. <https://doi.org/10.1038/s41580-022-00456-z>.
- Singh, H., Kumar, D., & Soni, V. (2022). Performance of chlorophyll a fluorescence parameters in *Lemna minor* under heavy metal stress induced by various concentration of copper. *Scientific Reports*, 12, 1-14. <https://doi.org/10.1038/s41598-022-14985-2>.
- Singh, A., & Satheeshkumar, P. K. (2024). Reactive Oxygen Species (ROS) and ROS Scavengers in Plant Abiotic Stress Response. In *Stress Biology in Photosynthetic Organisms: Molecular Insights and Cellular Responses* (pp. 41-63). Singapore: Springer Nature Singapore. https://doi.org/10.1007/978-981-97-1883-2_3
- Souza, L. R. R., Bernardes, L. E., Barbetta, M. F. S., & da Veiga, M. A. M. S. (2019). Iron oxide nanoparticle phytotoxicity to the aquatic plant *Lemna minor*: effect on reactive oxygen species (ROS) production and chlorophyll a/chlorophyll b ratio, *Environmental Science and Pollution Research*, 26(23), 24121-24131. <https://doi.org/10.1007/s11356-019-05713-x>.

- Stephenie, S., Chang, Y. P., Gnanasekaran, A., Esa, N. M., Gnanaraj, C. (2020). An insight on superoxide dismutase (SOD) from plants for mammalian health enhancement. *Journal of Functional Foods*, 68, 1-10. <https://doi.org/10.1016/j.jff.2020.103917>.
- Strzałka, K., Kostecka-Gugała, A., & Latowski, D. (2003). Carotenoids and environmental stress in plants: significance of carotenoid-mediated modulation of membrane physical properties. *Russian Journal of Plant Physiology*, 50(2), 168-173. <https://doi.org/10.1023/A:1022960828050>.
- Sun, S., Li, X., Sun, C., Cao, W., Hu, C., Zhao, Y., & Yang, A. (2019). Effects of ZnO nanoparticles on the toxicity of cadmium to duckweed *Lemna minor*. *Science of the Total Environment*, 662, 697-702. <https://doi.org/10.1016/j.scitotenv.2019.01.275>.
- Tamilselvam, K., Braidy, N., Manivasagam, T., Essa, M. M., Prasad, N. R., Karthikeyan, S., Thenmozhi, A. J. & Guillemin, G. J. (2013). Neuroprotective effects of hesperidin, a plant flavanone, on rotenone-induced oxidative stress and apoptosis in a cellular model for Parkinson's disease. *Oxidative medicine and cellular longevity*. 1, 1-11. <https://doi.org/10.1155/2013/102741>.
- Teixeira, G. C. M., de Mello Prado, R., Oliveira, K. S., D'Amico-Damião, V., & Junior, G. D. S. S. (2020). Silicon increases leaf chlorophyll content and iron nutritional efficiency and reduces iron deficiency in Sorghum plants. *Journal of Soil Science and Plant Nutrition*, 20, 1311-1320. <https://doi.org/10.1007/s42729-020-00214-0>.
- Wani, A. B., Chadar, H., Wani, A. H., Singh, S., & Upadhyay, N. (2017). Salicylic acid to decrease plant stress. *Environmental Chemistry Letters*, 15(1), 101-123. <https://doi.org/10.1007/s10311-016-0584-0>.
- Wang, Y., Cui, X., Yang, B., Xu, S., Wei, X., Zhao, P., Niu, F., Sun, M., Wang, C., Chieng, H., & Jiang, Y. Q. (2020). WRKY55 transcription factor positively regulates leaf senescence and the defense response by modulating the transcription of genes implicated in the biosynthesis of reactive oxygen species and salicylic acid in Arabidopsis. *Development, The Company of Biologists*, 147, 1-15. <https://doi.org/10.1242/dev.189647>.
- Yu, J., Cang, J., Lu, Q., Fan, B., Xu, Q., Li, W., & Wang, X. (2020). ABA enhanced cold tolerance of wheat 'dn1' via increasing ROS scavenging system. *Plant Signaling and Behavior*, 15(8), 1-11. <https://doi.org/10.1080/15592324.2020.1780403>.