

Evaluation of F_5 Individuals Obtained from B28×Kunduru-1149 Reciprocal Cross Population by Functional Markers

Gülan AYDEMİR¹, Ziya DUMLUPINAR^{24⁰,} İlker YÜCE³, Tuğba BAŞKONUŞ⁴, Selçuk SUNULU⁵ Hüseyin GÜNGÖR⁶

^{1,2,3}KSU Agricultural Biotechnology Department, Kahramanmaraş, Turkey, ^{4,5}KSU Field Crops Department, Kahramanmaraş, Turkey ⁶Duzce University Field Crops Department, Düzce, Turkey

¹https://orcid.org/0000-0001-6054-8893, ²https://orcid.org/0000-0003-3119-6926, ³https://orcid.org/0000-0002-9761-3561, ⁴https://orcid.org/0000-0002-0744-6086, ⁵https://orcid.org/0000-0002-1806-2547, ⁶https://orcid.org/0000-0001-6708-6337 ⊠: zdumlupinar@ksu.edu.tr

ABSTRACT

In the study, B28 and Kunduru-1149 durum wheat genotypes were crossed as reciprocal in 2012-2013 cropping season. 13 (B28/Kunduru-1149 and Kunduru-1149/B28) reciprocal crosses were obtained and were used as materials at F5 stage. The cross combinations and the parents were screened with 10 DNA markers to determine alleles of gluten strength (Bx7^{OE}), Yellow rust (Sun104, Xgwm18, Xwgp115 and Xgwm47), stem rust (Sun209 and Sun479), high protein ratio (UHW89), powdery mildew (Xgwm66) and leaf rust (Xgwm130). In the study, the average polymorphism information content (PIC) was calculated as 0.98 and the lowest PIC value was obtained from Xwgp115 marker with 0.96, while the rest of the markers had 0.99 PIC values. Stem rust resistance allele Sr49 was detected in B28/Kunduru-1149_F₅_4 (Sun479) and B28/Kunduru-1149_F₅_1 (Sun209) genotypes. One of the yellow rust resistance alleles Yr15(Xgwm18)was detected in B28/Kunduru-1149 F₅ 2 and B28/Kunduru-1149_ F_{5} 3 genotypes, while Yr51 (Sun104) was identified in B28/Kunduru-1149_F₅_3, B28/Kunduru-1149_F₅_6, B28/Kunduru-1149_F5_7, Kunduru-1149/B28_F5_2 and Kunduru-1149/B28_F5_6 genotypes. A dendrogram was created to determine kinship of the individuals with the parents. The highest genetic similarity was observed between B28 / Kunduru-1149 F_5 6 and Kunduru-1149 / B28_F5_2 genotypes with 0.714, while the most diverse ones were Kunduru-1149 and B28/Kunduru_F₅7 with 0.10.

Research Article

Article HistoryReceived: 11.02.2020Accepted: 24.02.2020

Keywords

Durum wheat Functional markers Yellow rust Stem rust Gluten strength

B28×Kunduru-1149 Resiproklu Melez Popülasyonundan Elde Edilen F₅ Bireylerinin Fonksiyonel Markörlerle Değerlendirilmesi

ÖZET

Bu çalışmada, B28 ve Kunduru-1149 makarnalık buğday genotipleri 2012-2013 ürün sezonunda resiproklu olarak melezlenmiş ve elde edilen 13 B28/Kunduru-1149-Kunduru-1149/B28 F5 kademesindeki hat materyal olarak kullanılmıştır. Melez kombinasyonları ve ebeveynler gluten dayanıklılığı (Bx70E), sarı pas (Sun104, Xgwm18, Xwgp115 ve Xgwm47), kara pas (Sun209 ve Sun479), yüksek protein oranı (UHW89), külleme (Xgwm66) ve kahverengi pas (Xgwm130) allelleri bakımından 10 DNA markörü ile taranmıştır. Çalışmada, ortalama polimorfizmi bilgi içeriği (PIC) 0.98 olarak hesaplanmış ve en düşük PIC değeri 0.96 ile Xwgp115 marköründen elde edilirken, diğer markörler 0.99 PIC değerine sahip olmuştur. B28/Kunduru- $1149_{F_5_4}$ (Sun479) ve B28/Kunduru-1149_F₅_1 (Sun 209)genotiplerinde kara pasa dayanıklılık alleli Sr49 tespit edilmiştir. Sarı pas dayanıklılık allellerinden Yr15 (Xgwm18) B28/Kunduru-1149_F5_2 ve B28/Kunduru-1149_F5_3 genotiplerinde bulunurken, Yr51 (Sun104) geni B28/Kunduru-1149_F5_3, B28/Kunduru-1149_F_5_6, B28/Kunduru-1149_F_5_7, Kunduru-1149/B28_F_5_2 ve Kunduru-1149/B28_F₅_6 genotiplerinde tanımlanmıştır. Ebeveynler

Araştırma Makalesi

Makale TarihçesiGeliş Tarihi÷ 11.02.2020Kabul Tarihi÷ 24.02.2020

Anahtar Kelimeler

Makarnalık buğday Fonksiyonel markörler Sarı pas Kara pas Gluten mukavemeti ile genotiplerin akrabalıklarını belirlemek için bir dendrogram oluşturulmuştur. En yüksek genetik benzerlik oranı B28 / Kunduru-1149_F₅_6 ve Kunduru-1149 / B28_F₅_2 genotipleri arasında 0.714 olarak tespit edilirken, birbirlerine en uzak genotipler 0.10 benzerlik oranı ile Kunduru-1149 ve B28/Kunduru_F₅_7 olmuştur.

To Cite : Aydemir G, Dumlupinar Z, Yüce I, Baskonus T, Sunulu S, Gungor H 2020. Evaluation of Individuals Obtained from B28×Kunduru-1149 Reciprocal Cross Population by Functional Markers. KSU J. Agric Nat 23 (4): 1005-1011. DOI: 10.18016/ksutarimdoga.vi.687935.

INTRODUCTION

Wheat (*Triticum* spp.) is an annual cereal crop consumed as major food source for centuries (Sevinç, 2010). Turkey is one of the centers of origin of the durum wheat which is originated in Karacadağ location of South-East Anatolia Region (Anonymous, 2016).

Wheat is grown in 219.3 mil. ha and 757.7 mil. ton produced world-wide, while 7.2 mil. ha grown and 20 mil. ton produced in Turkey (TUIK, 2018). World durum wheat production is approximately 40.2 mil. ton, while 3.6 mil. ton in Turkey (TUIK, 2018). Turkey is one of the durum wheat producers in the world especially in South Anatolia Region.

Durum wheat (2n=4x=28, AABB) differs for its utilization from bread wheat (2n=6x=42, AABBDD) as pasta, bulgur and couscous were made from durum wheat, while bread, noodle etc. were made from hexaploid bread wheat.

Plant breeders have put effort on developing high quality and high yielding cultivars for many years and different breeding techniques are used to improve crop-plants. Crossing is one of the breeding techniques used to combine traits from both parents.

Landraces have been sources to expand genetic diversity for disease resistance, drought tolerance, quality traits and many more traits. Besides favorable characteristics, they have some negative traits such as lodging, low grain yield etc. which must be eliminated by improving the traits.

Molecular marker technology is used widely in breeding programs with recent developments in biotechnology. Markers assisted selection (MAS) has been used to detect alleles related to the traits such as disease, quality parameters and agronomic traits. It provides earlier and precise selection especially for quantitative traits. Functional markers have been developed after many efforts on quantitative trait locus (QTL) studies and available for many genes and traits. Marker assisted selection have now been used widely due to its accurate, rapid, reproducible and cost effective solutions for the breeding programs.

Kunduru-1149 is a cultivar developed from selection of landraces. It has many advantages besides disadvantages such as lodging and lower quality parameters. On the other hand B28 is a landrace obtained from USDA National Small Grains Collection, Aberdeen, USA gene bank and has higher quality traits and disease resistance. Those genotypes crossed as reciprocal and 13 genotypes at F₅ stages selected from both combinations (B28 × Kunduru-1149 and Kunduru-1149 × B28). In the study it was aimed to identify some disease and quality traits by allele specific markers to determine genotypes related with those gluten strength, high protein ratio, yellow rust, stem rust, powdery mildew and leaf rust diseases genes. For this purpose, the genotypes were screened with 10 allele specific DNA markers.

MATERIALS and METHOD

In the study, 13 genotypes obtained from reciprocal crosses of B28 and Kunduru-1149 and the parents were used as plant material. Seeds of the genotypes were planted and two leaves seedling were harvested to extract DNAs (Dumlupinar, 2016). The DNA content and purity were determined by spectrophotometer (Thermo-Scientific Nanodrop 2000 spectrophotometer).

Allele specific markers of gluten strength (Bx7^{oE}), Yellow rust (Sun104, Xgwm18, Xwgp115 and Xgwm47), stem rust (Sun209 and Sun479), high protein ratio (UHW89), powdery mildew (Xgwm66) and leaf rust (Xgwm130) were screened on Qiagen Qiaxcel Fragment Analyzer (Table 1). The data obtained from fragment analyzer were scored and the genetic similarity of the genotypes were determined by Dice index (Dice, 1945) using NTSYSpc 2.21q software (Rohlf, 2005). Polymorphism information content (PIC) was determined by using the formula described by Weir (1996), where PIC=1- Σ Pi², where P_i is the frequency of the *i*th allele in the 15 durum wheat genotypes studied.

RESULTS and DISCUSSION

Marker assisted selection (MAS) studies in wheat have been accomplished in many plant breeding programs worldwide and succeeded on many agronomical traits. Based on genotypic data obtained from durum wheat segregation populations was investigated. The allele numbers of the primers, figures of the primers Xgwm18 and Xgwm66 were shown in Figures 1, 2 and 3 respectively. The dendrogram was generated from the marker data of durum wheat genotypes is shown in Fig. 4. The allele specific markers interrogated on the durum wheat genotypes were indicated in Table 2.

Table 1 DNA Primers Used in the Study	
Çizelge 1 Çalışmada kullanılan DNA Primerler	i

No (<i>Numara</i>)	Primer Name (Primer Adı)	Primer Sequence(5·3') (Primer Dizisi (5·3'))	Reference (Referans)	Loci (Lokus)	Expected Frqagment Size (bp) (Beklenen Bant Ulzunuluğu (hc))	Marker Type (Markör Tipi)	
1	$egin{array}{c} Bx7^{OE}F\\ Bx7^{OE}R\end{array}$	CCTCAGCATGCAAACATGCAGC CTGAAACCTTTGGCCAGTCATGT C	Butow et al., 2003	Glutent Strength	563	Co- dominant	
2	SUN104_F SUN104_R	TGCTATGTGCGTGATGATGA TTACATGCTCCAGCGACTTG	Randhawa et al., 2014	Yellow Rust <i>Yr51</i>	225	Dominant	
3	SUN209_F SUN209_R	AG CTATGAGCTTCGCTATTG GTGATTGGTTCGGATTACTTA	Bansal et al., 2015	Stem Rust Sr49	148	Co- dominant	
4	SUN479_F SUN479_R	CAAATGAAATGTGATCCTGTT TCATCTAACCAGCAATGGTAT	Bansal et al., 2015	Stem Rust Sr49	200	Co- dominant	
5	UHW89_BF UHW89_R	TCTCCAAGAGGGGGAGAGACA TTCCTCTACCCATGAATCTAGCA	Distelfeld et al., 2006	High Protein Content <i>Gpc-B1</i>	122	Co- dominant	
6	XGWM18_F XGWM18_R	TTGCTACCATGCATGACCAT TTCACCTCGATTGAGGTCCT	rgctaccatgcatgaccat Roder et al., 1998 Yellow rcacctcgattgaggtcct Roder et al., 1998 Yust yr15 and 186, 190				
7	XGWM47_F XGWM47_R	TTGCTACCATGCATGACCAT TTCACCTCGATTGAGGTCCT	Roder et al., 1998	Yellow Rust <i>Yr64</i> and <i>Yr66</i>	190	Co- dominant	
8	XGWM66_F XGWM66_R	TTGCTACCATGCATGACCAT TTCACCTCGATTGAGGTCCT	Roder et al., 1998	Powdery Mildew	137	Co- dominant	
9	XGWM130_F XGWM130_R	AGCTCTGCTTCACGAGGAAG CTCCTCTTTATATCGCGTCCC	Roder et al., 1998	Leaf Rust <i>Lr34</i>	121,126	Co- dominant	
10	XWGP115_F XWGP115_R	AGTGTCTTGTAGGGTATC TCAGGCCGTGAAAAATAT	Roder et al., 1998	Yellow Rust <i>Yr45</i>	492	Co- dominant	



Figure 1 Allele numbers of the primers screened for the durum wheat reciprocal segregation populations Şekil 1 Makarnalık buğday resiprokal melez popülasyonunda görüntülenen primerlerin allel sayıları

5000																	- 3000	
Peak Size	500 —																	Peak Size
250	300 — 200 —																	
150 75 25	100 — 50 —																182	
		Kunduru-1149	B28	B28/Kunduru-1149_F ₅ _1	$B28/Kunduru-1149_F_{5_2}$	B28/Kunduru-1149_F ₅ _3	$B28/Kunduru-1149_F_{5_4}$	$B28/Kunduru-1149_F_{5}5$	$B28/Kunduru-1149_F_{5_6}$	$B28/Kunduru-1149_F_{5_}7$	Kunduru-1149/B28_F ₅ _1	Kunduru-1149/B28_F ₅ _2	Kunduru-1149×B28_F ₅ _3	Kunduru-1149/B28_F _{5_} 4	Kunduru-1149/B28_F ₅ _5	Kunduru-1149/B28_F ₅ _6	10	

Figure 2 Visualization of Xgwm18 (Yr26) DNA marker which produced alleles on B28/Kunduru-1149_F₅_2 and B28/Kunduru-1149_F₅_3 genotypes on Fragment Analyzer

Şekil 2 Fragment analiz cihazında, B28 / Kunduru-1149_F5_2 ve B28 / Kunduru-1149_F5_3 genotiplerinde alleller üreten Xgwm18 (Yr26) DNA markörünün görüntülenmesi



Figure 3 Visualization of Xgwm66 DNA marker which could not produce powdery mildew resistance gene along the durum wheat genotypes on Fragment Analyzer

Şekil 3 Fragment Analiz cihazında makarnalık buğday genotiplerinde küllemeye dayanıklılık genini üretemeyen Xgwm66 DNA markörünün görüntülenmesi



Figure 4 A dendogram was created using genetic similarity index of the durum wheat genotypes Şekil 4 Makarnalık buğday genotiplerinin genetik benzerlik indeksi kullanılarak oluşturulmuş filogenetik ağaç

Table 2 Allelic variation of B28, Kunduru-1149, Kunduru-1149/B28 and B28/Kunduru-1149 genotypes *Çizelge 2 B28, Kunduru-1149, Kunduru-1149/B28 ve B28/Kunduru-1149 melez kombinasyonlarınun allellik varyasyonları*

Markers (Markörler)		68	60		62	115	418	1130	166	147
Genotypes (Genotipler)	SUN 1	3MHU	SUN2(BX70E	SUN4′	XWGP	XGWJ	XGWI	XGWJ	XGWI
Kunduru-1149										
<i>B28</i>										
<i>B28/Kunduru-1149_F5_1</i>			+							
<i>B28/Kunduru-1149_F5_2</i>							+			
<i>B28/Kunduru-1149_F5_3</i>	+						+			
<i>B28/Kunduru-1149_F5_4</i>					+					
<i>B28/Kunduru-1149_F5_5</i>										
<i>B28/Kunduru-1149_F5_6</i>	+									
<i>B28/Kunduru-1149_F₅_7</i>	+									
Kunduru-1149/B28_F ₅ _1										
Kunduru-1149/B28_F ₅ _2	+									
Kunduru-1149/B28_F ₅ _3										
Kunduru-1149/B28_F ₅ _4										
Kunduru-1149/B28_F5_5										
Kunduru-1149/B28_F5_6	+									
PIC Values (%)	99	99	99	99	99	96	99	99	99	99

In terms of expected primer sizes, the allele specific markers data evaluated on segregation durum wheat populations.

In the study the functional markers for gluten strength (Bx7OE), Yellow rust (Sun104, Xgwm18, Xwgp115 and Xgwm47), stem rust (Sun209 and Sun479), high protein ratio (UHW89), powdery mildew (Xgwm66) and leaf rust (Xgwm130) were used. According to the results a total number of 50 alleles were produced by 10 DNA primers and, average allele number per

primer was 5. The average polymorphism information content of the study was determined as 98%, and the lowest PIC value was obtained from Xwgp115 marker with 96%, while the rest of the primers had a 99% PIC values (Table 2). In a previous study Maccaferri et al. (2003) conducted a research on genetic diversity of a durum wheat set derived from Mediterranean basin using microsatellites and reported a mean diversity index (DI) of 56%. Moragues et al. (2007) investigated the genetic variation of 63 durum wheat landraces and indicated an average PIC value of 24% for AFLP and 70% for SSR markers. Also Gungor (2019) reported an average PIC value of 72.5% on a durum wheat cultivar panel derived from different breeding programs.

B28/Kunduru-1149_F₅_3, B28/Kunduru-1149_F5_6, B28/Kunduru-1149 F5 7, Kunduru-1149/B28 F5 2 and Kunduru-1149/B28_F5_6 genotypes carried alleles for yellow rust resistance gene Yr51 as reported and expected size of 225 bp (Randhawa et al., 2014). Gungor (2019) also detected 225 bp alleles using the Sun104 marker for yellow rust on a durum wheat panel. Yan et al. (2003) reported a marker-trait relation for Sun104 marker and yellow rust resistance gene Yr51. In addition, B28/Kunduru-1149_F5_2 and B28/Kunduru-1149_F5_3 genotypes had alleles for Xgwm18 markers related with yellow rust resistance gene Yr15 and Yr26. However, the other markers related with yellow rust resistance such as Xwgp115 and Xgwm47 had no alleles among the genotypes, though Cowger et al., 2012) reported a marker-trait relation for Xgwm47 for Yr64 and Yr66 yellow rust resistance and Gungor (2019) indicated an allele for Xwgp115 marker in durum wheat cultivars (Table 2). Of the Sun209 and Sun479 markers linked to stem rust resistance gene Sr49, Sun209 had allele on B28/Kunduru-1149_F₅1 genotype at 148bp, while Sun479 amplified allele on B28/Kunduru-1149_F5_4 genotype at 200 bp. Bansal et al. (2015) and Gungor (2019) reported relation for the Sun209 and Sun479 markers with Sr49 stem rust gene at 148 bp and 200 bp respectively, which is consistent with our findings.

On the other hand, some of the markers such as Xgwm130 marker linked to the leaf rust resistance gene, $Bx7^{OE}$ marker for gluten strength, UHW89 marker which is carrying high protein content gene *Gpc-B1* and Xgwm66 marker which is involved with powdery mildew resistance gene used in the study had no alleles on the genotypes. However, Butow *et al.* (2003), Cho *et al.* (2017), Liang *et al.* (2010) and Gungor (2019) indicated that Bx7OE marker produced allele on their genotype panel and related with gluten strength and improved dough strength. In addition, Distelfeld *et al.* (2006) indicated marker-trait relation for UHW89 and high protein content, consistent with Gungor (2019).

A dendrogram was created using the whole alleles produced by DNA markers. According to the dendrogram two main groups obtained. The first one was consisted of B28/Kunduru-1149_F₅_1, B28/Kunduru-1149_F₅_2, B28/Kunduru-1149_F₅_3 and Kunduru-1149/B28_F5_3 genotypes, while the parents were took place on the other group. The second group divided into two groups including parents in each group. The most similar genotypes were found as Kunduru-1149/B28_F₅_2 and B28/Kunduru- $1149_{F_{5}}$ genotypes with the 71% genetic similarity (Figure 4). Ren et al. (2013) indicated a narrow genetic base on a worldwide germplasm accession of durum wheat released in 1960s and 1970s, though a rapid increase after 1970s. Gungor (2019) reported a broad genetic variation in durum wheat cultivars derived from different collections and breeding programs.

CONCLUSION

Two reciprocal cross combinations of B28/Kunduru-1149 and Kunduru-1149/B28 were screened for the 10 allele specific DNA markers. It is determined that B28/Kunduru-1149_F₅_4 B28/Kunduruand 1149_F₅1 genotypes had stem rust resistance allele Sr49. B28/Kunduru-1149_F₅_2 and B28/Kunduru-1149_F₅_3 genotypes had yellow rust resistance alleles Yr15, while yellow rust resistance allele Yr51 was detected on B28/Kunduru-1149 F5 3. B28/Kunduru-1149 F₅ 6, B28/Kunduru-1149_F₅7, Kunduru- $1149/B28_{F_5}2$ Kunduru-1149/B28_F5_6 and genotypes. On the other hand, according to the dendrogram B28 / Kunduru-1149_F5_6 and Kunduru-1149 / $B28_{F_5_2}$ genotypes were found the most similar genotypes with 0.714, while Kunduru-1149 and B28/Kunduru_F5_7 with 0.10 combinations were found the most diverse ones.

ACKNOWLEDGEMENT

This study is produced from the master thesis of Gülan AYDEMİR. The other authors were responsible for making crosses and bringing generations from F_1 to F_5 . The abstract is also published in International Biological, Agricultural & Life Science Congress November 7-8, 2019 Lviv, Ukraine.

Authors also would like to thank The Scientific Research Projects Council of Kahramanmaraş Sütçü İmam University (Project Number 2017/7-175 YLS).

Statement of Conflict of Interest

Authors have declared no conflict of interest.

Author's Contributions

The contribution of the authors is equal.

REFERENCES

- Anonymous 2016. Turkey's Wheat Atlas, © WWF-Turkey (Wildlife Protection Foundation). http://d2hawiim0tjbd8.cloudfront.net/downloads/tu rkiye_nin_buday_atlas_web.pdf Date of Access: 19.05.2019
- Bansal UK, Muhammad S, Forrest KL, Hayden MJ, Bariana HS 2015. Mapping of a New Stem Rust Resistance Gene Sr49 in Chromosome 5B of Wheat. Theor Appl Genet 128 (10): 2113-2119.
- Butow BJ, Ma W, Gale KR, Cornish GB, Rampling L, Larroqueand O, Bekes F 2003. Molecular Discrimination of Bx7^{OE} Alleles Demonstrates That a Highly Expressed high-Molecular-Weight Glutenin Allele Has a Major Impact on Wheat

Flourdough Strength. Theor Appl Genet 107(8), 1524–1532.

- Cho SW, Roy SK, Chun JB, Cho K, Park CS 2017. Overexpression of The Bx7 High Molecular Weight Glutenin Subunit on The Glu-B1 Locus in a Korean Wheat Landrace. Plant Biotechnol. Rep 11: 97–105.
- Cowger C, Miranda L, Griffey C, Hall M, Murphrand JP, Maxwells J 2012. Wheat Powdery Mildew, Disease Resistance in Wheat. Sharma, I. (ed.) CABI Publications London, UK.
- Dice LR 1945. Measures of The Amount of Ecologic Association Between Species. Ecology 26: 297–302.
- Distelfeld A, Uauy C, Fahimaand T, Dubcovsky J 2006. Physical Map of The Wheathigh-Grain Protein Content Gene Gpc-B1 and Development of a High-Through Put Molecular Marker. New Phytologist 169: 753–763.
- Dumlupinar Z, Brown R, Campbell R, Jellen EN.
 Anderson J, Bonman M, Carson M, Chao S, Obert D, Jackson E 2016. The Art of Attrition:
 Development of Robust Oat Microsatellites. Plant Breed 135: 323–334.
- Gungor H 2019. Allelic Variations and Agronomic Comparisons Of Durum Wheat Cultivars Under East-Mediterranean Conditions. Intl. J. Agric. Biol 21: 891–898.
- Liang D, Tang J, Pena RJ, Singh R, He X, Shen X, He Z 2010. Characterization of CIMMYT Bread Wheats for High-And Low-Molecular Weight Glutenin Subunits and Other Quality-Related Genes with SDS-PAGE, RP-HPLC and Molecular Markers. Euphytica 172: 235-250.
- Maccaferri M, Sanguineti MC, Donini P, Tuberosa R 2003. Microsatellite Analysis Reveals a Progressive Widening of The Genetic Basis in The Elite Durum Wheat Germplasm. Theor Appl Genet 107: 783-797.

Moragues M, Moralejo M, Sorrells ME, Royo C 2007.

Dispersal of Durum Wheat [*Triticum Turgidum* L. Ssp. Turgidum Convar. Durum (Desf.) MacKey] Landraces Across The Mediterranean Basin Assessed by AFLPs and Microsatellites. Genet Resour Crop Evol 54: 1133-1144.

- Randhawa M, Bansal U, Valarik M, Klocova B, Dolezeland J, Bariana H 2014. Molecular Mapping Of Stripe Rust Resistance Gene Yr51 in Chromosome 4 AL of Wheat. Theor Appl Genet 127: 317-324.
- Ren J, Sun D, Chen L, You FM, Wang J, Peng Y, Nevo E, Sun D, Luo MJ, Peng J 2013. Genetic Diversity Revealed by Single Nucleotide Polymorphism Markers in a Worldwide Germplasm Collection of Durum Wheat. Intl J Mol Sci 14: 7061-7088.
- Roder MS, KorzunV, Wendehake K, Plaschke J, Tixier M, Leroyand P, Ganal MW 1998. A Microsatellite Map Of Wheat. Genetics, 149: 2007-2023.
- Rohlf FJ 2005. NTSYS-pc: Numerical Taxonomy and Multivariate Analysis System Version 2.2. Setauket, Exeter Publishing, NY, USA.
- Sevinç D 2010. Sarı Pas Hastalığına Dayanıklı ve Duyarlı Buğday (Triticum aestivum L.) Çeşitlerinin Moleküler Markörler ile İncelenmesi. İstanbul Üniversitesi Fen Bilimleri Enstitüsü Biyoloji Anabilim Dalı Botanik Programı, Yüksek Lisans Tezi, 102s.
- TUIK 2018. Turkish Statistical Institute. www.tuik.gov.tr. Date of Access 05.06.2019.
- Weir BS 1996. Genetic Data Analysis II, 2nd ed. Sinauer Associates Inc. ISBN-13: 978-0878939022 Sunderland, Massachusetts, USA. 437s.
- Yan GP, Chen XM, Line RF, Wellings CR 2003. Resistance Gene-Analog Polymorphism Markers Co-Segregating with The YR5 Gene for Resistance to Wheat Stripe Rust. Theor Appl Genet 106: 636– 643.