

Anticancer, Antiproliferative and Lactate Dehydrogenase Enzyme Activities of Astragalus elongatus subsp. nucleiferus on Human Cancer Cells

Nazım SEKEROGLU¹⁴⁷, Sevgi GEZİCİ², Ceren Serin TANRIOVER³, Fatih YAYLA⁴

¹Department of Food Engineering, Faculty of Engineering and Architecture, Kilis 7 Aralık University, Kilis, ^{2,3}Department of Molecular Biology and Genetics, Faculty of Science and Literature, Kilis 7 Aralik University, Kilis, ⁴Department of Biology, Faculty of Science and Literature, Gaziantep University, Gaziantep

¹https://orcid.org/0000⁻0002⁻0630⁻0106, ²https://orcid.org/0000⁻0002⁻4856⁻0221, ³https://orcid.org/0000⁻0002⁻4974⁻6669, ⁴https://orcid.org/0000⁻0002⁻6490⁻6288,

⊠: sekeroglu@kilis.edu.tr

ABSTRACT

In this study, potential anticancer and antiproliferative activities of ethanol and water extracts from aerial parts and roots of Astragallus elongatus subsp. nucleiferus were evaluated. MTT, trypan blue, and LDH enzyme activity assays were performed to determine cytotoxicity and cell proliferation potentials of the plant extracts against human A549, H1299, C6 cancer cells, and non-tumorous HUVECs. As results of MTT and trypan blue assays, dose-dependent anticancer and antiproliferative effects were observed on tested human lung and brain carcinoma cells. The water extracts obtained from the root exerted higher anticancer activity with IC50 values ranging from 5.81±0.46 to 18.24±0.12µg/mL than the extracts of aerial parts. In contrary anticancer and cytotoxicity results, the ethanol extract of the root was also caused to the highest apoptosis level in a dose dependently. Regarding LDH activity, the plant extracts obtained from aerial parts and roots were demonstrated the highest LDH activity towards H1299 cells. The obtained results showed that the roots of the plant are able to inhibit cell growth in cancer cells in a time and concentration dependently.

Research Article

Article History	у
Received	26.03.2019
Accepted	: 13.06.2019

Keywords

Anticancer, Apoptosis, *Astragalus* sp., Cytotoxicity, Cell proliferation

İnsan Kanser Hücrelerinde *Astragalus elongatus* subsp. *nucleiferus*' un Antikanser, Antiproliferatif ve Laktat Dehidrojenaz Enzim Aktiviteleri

ÖZET

Bu araştırmada, Astragallus elongatus subsp. nucleiferus' un toprak üstü ve kök kısımlarından elde edilen etanol ve su özütlerinin potansiyel antikanser ve antiproliferatif aktivitelerinin araştırılması amaçlanmıştır. Bitki özütlerinin A549, H1299, C6 insan kanser hücreleri ve kanserli olmayan HUVEC hücrelerine karşı sitotoksisite ve hücre proliferasyon potansiyellerini araştırmak için; MTT, tripan mavisi ve LDH enzim aktivite analizleri yapılmıştır. MTT ve tripan mavisi deneylerinin bir sonucu olarak, test edilen insan akciğer ve bevin karsinoma hücrelerinde doza bağlı antikanser ve antiproliferatif etkiler gözlenmiştir. Bitkinin kökünden elde edilen su özütleri, 5.81±0.46 ile 18.24±0.12µg / mL arasında değişen IC₅₀ değerleriyle, toprak üstü özütlerinden daha yüksek bir antikanser aktivite göstermiştir. Antikanser ve sitotoksisite sonuçlarının aksine, kök etanol özütleri doza bağlı olarak en yüksek apoptoz seviyesine neden olmuştur. LDH aktivitesi ile ilgili olarak; toprak üstü kısımdan ve köklerden elde edilen bitki özütlerinin H1299 hücrelerine karşı en yüksek LDH aktivitesi göstermiştir. Elde edilen sonuçlar; bitki köklerinin, kanser hücrelerinde hücre büyümesini konsantrasyon ve zamana bağlı olarak önleyebildiğini göstermiştir.

Araştırma Makalesi

Makale Tarihçesi Geliş Tarihi ÷ 26.03.2019 Kabul Tarihi ÷ 13.06.2019

Anahtar Kelimeler Antikanser, Apoptoz, *Astragalus* sp., Sitotoksisite, Hücre proliferasyonu

To Cite: Sekeroglu, N, Gezici S, Tanrıover CS, Yayla F 2019. Anticancer, Antiproliferative and Lactate Dehydrogenase Enzyme Activities of *Astragalus elongatus* subsp. *nucleiferus* on Human Cancer Cells. KSU J. Agric Nat 22(Suppl 1): 24-29. DOI: 10.18016/ksutarimdoga.v22i49073.569493.

INTRODUCTION

In recent years, there has been a widespread interest in herbal medicines due to their therapeutic and prevention potentials in cancer cases. Boosting the immune system is the most effective remedy strategy in most cancers (Newman and Cragg, 2016; Roleira et al., 2018; Gezici and Sekeroglu, 2019a). From this point of view, immune stimulating plants have critically importance for cancer patients. The genus *Astragalus* L., the largest in the Fabaceae family, are of important medicinal plants that known as immunostimulant, hepatoprotective, antiviral, antidepressive, and anti-inflammatory agents (Block and Mead, 2003; Ionkova et al., 2014).

The plants of Astragalus genus are rich of bioactive secondary metabolites such as polysaccharides, saponins, and cycloartanes that can deeply affect to immune system (Shao et al., 2004; Yalçın et al., 2012; Ionkova et al., 2014). Their wide range of bioactive metabolites are responsible for their diverse pharmacological effects, leading us to investigate biological activities of wild grown Astragalus species in Turkey, particularly under mild-climate areas in sandy soils. In our previous study, aerial parts and root extracts of A. elongatus subsp. nucleiferus were analyzed for their potential neuroprotective effects and antioxidant capacities along with polyphenolic contents (Gezici et al., 2018). Taking our previous results, we aimed to screen potential anticancer and antiproliferative activities of extracts from aerial parts and roots of A. elongatus subsp. nucleiferus in the presented research.

Although there is scientific evidence about therapeutic properties of Astragalus species, there has been no report yet determining anticancer potentials of aerial parts and root extracts from A. elongatus subsp. nucleiferus against human lung carcinoma (A549), non-small lung cancer (H1299), and glioma (C6) cancer cells. According to literature survey, our findings in this paper ought to be the first report, which was carried out for investigation of *in vitro* anticancer, antiproliferative, and lactate dehydrogenase releasing activities of water and ethanol extracts obtained from aerial part and root of A. elongatus subsp. nucleiferus.

MATERIALS AND METHODS

Plant Material

The plant samples were collected during the months of April-May 2017 from Gaziantep, Turkey. Taxonomic identification of the plant was performed by a senior taxonomist Fatih Yayla, from Department of Biology, Gaziantep University (Gaziantep, Turkey). The voucher specimen was deposited at the Herbarium of Department of Biology, Gaziantep University, Turkey (GAUN1706). The plant materials were dried in shade and stored at 25°C in dark until extraction.

Preparation of Crude Extracts

The air-dried and powdered plant material (50 g) obtained from aerial part and root of *A. elongatus* subsp. *nucleiferus* were extracted with distilled water (dH₂O) and ethanol (EtOH) by maceration extraction procedure as described in previous study (Gezici and Sekeroglu, 2019b). Extract yields of the EtOH and dH₂O of the aerial parts and roots were 10.606%, 7.198%, 8.926%, and 5.312% (w/w), respectively.

Cell Lines and Culture Conditions

Human lung and human brain cancer cell lines: A549 (lung carcinoma), H1299 (non-small cell lung cancer), C6 (glioma) and non-tumorous HUVECs (human umbilical vein endothelial cells), purchased from the American Type Culture Collection (ATCC, USA) for analysis. A549 and H1299 lung cancer cells were cultured in Roswell Park Memorial Institute Medium (RPMI, ThermoFisher Scientific), and the other cell lines were grown in Dulbecco's modified Eagle medium (DMEM): Ham's F12 nutrient medium (1:1) (ThermoFisher Scientific) in the flasks at 37°C in a humidified CO_2 (5%) incubator. All the media and laboratory conditions were the same as given in our previous studies (Gezici et al., 2017; Gezici, 2018; Gezici, 2019a). All the experiments were repeated minimum of three times.

Anticancer Activity Determination by MTT Assay

MTT (3-(4,5-dimethylthiazol-2-yl)-2,5diphenyltetrazolium bromide) assay was used to determine cytotoxic properties of the various extracts according to the method of Mosmann (1983) with small changes. Briefly, densities of $5 \ge 10^3$ cells were seeded in 200 µl medium into 96-well plates for 24 h, and A. elongatus subsp. nucleiferus extracts with different concentrations ranged from 10 to 200 µg/mL were added to wells, and then all the wells incubated at 37°C for 48 h. The medium was discarded and 50 µL/well of MTT solution (5 mg/mL) was added into each well after incubation. And then, the absorbance was measured at 570 nm with a Thermo Lab systems 408 Multiskan multiplate spectrophotometer, and the dose response curve was used to generate the IC_{50} (µg/mL) values for each cell line.

Antiproliferative Activity Determination by Trypan Blue Assay

Extracts' antiproliferative activities were evaluated with Trypan Blue exclusion method described by Strober (2001) for A549, H1299, C6, and HUVEC cell lines. Chemicals and conditions used in the all analysis were the same as in our former publications (Gezici et al., 2017; Gezici, 2019b). Stock concentration as 1 mg/mL for all the *A. elongatus* subsp. *nucleiferus* extracts were prepared in DMSO and used at different concentrations (10, 50, 100, and 200 μ g/mL) from each extract, and they were incubated at 37°C for 24, 48, and 72 hours. The cell viability was determined by microscopically (Nikon, Japan), and the viable cells were counted by Cedex XS Analyzer, an automated cell counter.

Lactate Dehydrogenase (LDH) Release Activity Determination

Release of the Enzyme Lactate Dehydrogenase (LDH) Activity assays were achieved by detection of necrosis in the cells according to method of Al-Qubaisi et al. (2011). Chemicals and conditions used for all the analysis were the same as given our former papers (Gezici, 2018; Gezici, 2019a). LDH percentage release in medium was calculated by comparing to total LDH in in the same well. In order to generate the IC₅₀ (μ g/mL) values, dose response curve was used.

Statistics

The data obtained from analysis were expressed as mean value and standard deviation of the mean (mean \pm SD) from at least triplicate analyses. A linear regression analysis was performed to calculate IC₅₀ values. P<0.05 and p<0.01 values were considered as statistically significant, and very significant, respectively.

RESULTS AND DISCUSSION

Anticancer Activity Results

MTT assay was assessed to evaluate the cytotoxic activities of the *A. elongatus* subsp. *nucleiferus* extracts on three human cancer cell lines (A549, H1299, and C6), compared to cytotoxic activity of non-tumorous HUVECs. Anticancer activity results are given in Table 1 regarding of IC_{50} values after 48h treatment period.

MTT assay results showed that the extracts of A. elongatus subsp. nucleiferus exhibited remarkable cytotoxicity towards the tested human lung and brain carcinoma cells even at lower dose and minimum exposure time. As summarized in the Table 1, the water extracts obtained from root was exerted higher anticancer activity on A549, H1299, and C6 calls, with the IC_{50} values 6.14 ± 0.02 , 5.81 ± 0.46 , and $10.68\pm0.19\mu$ g/mL (at 200 μ g/mL concentration p<0.01), respectively, than the extracts obtained from the aerial parts of the plant. Furthermore, the ethanol extract obtained from aerial part of the plant was exerted lower cytotoxicity towards the cancer cells, compared to that of the water extract as dose and time dependently. In terms of the cancer cell types, all the extracts were found to be highly cytotoxic with IC50 values in range of 5.81±0.46 - 10.75±1.16µg/mL on H1299 cells at the highest concentration for 48h, which was followed by A549 and C6 cells, respectively (Table 1). Based on the findings on cytotoxicity, *A. elongatus* subsp. *nucleiferus* with aerial part and root demonstrated anticancer activity that against the cancer cells in a dose dependent manner, thus the plant could have significant potential in the prevention of cancer development.

Antiproliferative Activity Results

To screen antiproliferative activity of the ethanol and water extracts of *A. elongatus* subsp. *nucleiferus*, trypan blue assay was used, and the cells were counted according to their viabilities after treated with different doses (10, 50, 100 and 200 μ g/mL) for 24, 48, 72h. Results of cell viability (%) percentage of A549, H1299, and C6 cancer cells by comparing with non-tumorous HUVECs were determined after 48h treatment at the highest concentration of the extracts (Figure 1).

In consistent with MTT assay, results of trypan blue assay demonstrated that cell growth and viability in the cancer cells were inhibited by the extracts from A. elongatus subsp. nucleiferus, in a time and dose dependent manner. As can be seen in the Figure 1, antiproliferative activity assay was also resulted in the superiority of the water extracts obtained from the root of the plant against H1299 lung carcinoma cells. On the other hand, water extracts of root showed significant reduction in cell growth of A549, H1299, and C6 cancer cells with survival percentage of 28%, 20%, and 35% against the cells, respectively. However, ethanol extract obtained from aerial part of the plant demonstrated lower reduction in cell growth with 46%, 40%, and 60% viability percentage to the A549, H1299, and C6 cancer cells, respectively, even at the lowest concentration (Fig 1.). The results revealed that A. elongatus subsp. nucleiferus extracts had quite inhibitory effects against to the cell growth in all the cancer cells.

According to the results we obtained from this research, the A. elongatus subsp. nucleiferus extracts inhibited the cell growth in lung carcinoma and brain glioma cancer cells in a dose-dependent manner. It was clear that increasing concentration of the extracts resulted in decreasing the number of live cells. Moreover, increasing exposure time was resulted in a significant inhibition of cell growth in the cancer cells. It is noteworthy that A. elongatus subsp. nucleiferus could have significant potential as an anticancer agent in the management of decreasing the amount of cancer cells. The anticancer and growth inhibitory effects of the extracts may be due to the fact that rich secondary metabolites components of the plant as described previously (Block and Mead, 2003; Shao et al., 2004; Ionkova et al., 2014; Ranjbar and Mahmoudian, 2015).



Figure 1. Cell viability (%) of A549, H1299 and C6 cancer cells

* Cells were treated with 200µg/mL concentration of the extracts for 48h.

 ** HUVEC cells were used as control and set as 100% cell viability

Table 1. Anticancer effects of <i>A. elongatus</i> subsp	. <i>nucleiferus</i> extracts against A549, H1299 and C6 cells for 48h

Cell Lines	s Plant Part-Extract Type		Concentrations of the Extracts (µg/mL) ^a			
Cell Lines			10µg/mL	50μg/mL	100µg/mL	200µg/mL
A549	Aerial Part	Ethanol	$58.67 \pm 1.02^{**}$	$46.36 \pm 0.38^{**}$	$43.85 \pm 0.70^{*}$	$13.07 \pm 0.64^{**}$
		Water	$51.45 \pm 0.96^{*}$	$40.91 \pm 1.06^{*}$	$32.03 \pm 0.08^{**}$	$10.08 \pm 1.14^{**}$
	Root Part	Ethanol	$46.09 \pm 0.01^{**}$	$36.04 \pm 1.54^{**}$	$24.79 \pm 0.68^{**}$	$8.15 \pm 0.18^{*}$
		Water	$48.01 \pm 0.18^{**}$	$30.96 \pm 0.15^{**}$	$22.25 \pm 1.02^{*}$	$6.14 \pm 0.02^{**}$
H1299	Aerial Part	Ethanol	$55.46 \pm 1.14^{**}$	$42.44 \pm 1.60^{*}$	$24.22 \pm 1.09^{**}$	10.75 ± 1.16
		Water	47.16 ± 1.26	$35.07 \pm 0.05^{**}$	$26.64 \pm 0.93^{*}$	$7.86 \pm 0.14^{**}$
	Root Part	Ethanol	$44.68 \pm 0.16^{**}$	$29.18 \pm 0.98^{*}$	$18.72 \pm 0.42^{**}$	$7.16 \pm 1.01^{**}$
		Water	$41.72 \pm 0.49^{**}$	$25.45 \pm 0.02^{**}$	$14.01 \pm 0.08^{**}$	$5.81 \pm 0.46^{**}$
C6	Aerial Part	Ethanol	$74.04 \pm 0.61^{**}$	$54.09 \pm 0.74^{**}$	$49.80 \pm 0.17^{**}$	$18.24 \pm 0.12^{**}$
		Water	$60.02 \pm 0.08^{*}$	$45.88 \pm 0.68^{*}$	$28.04 \pm 0.96^{**}$	$12.69 \pm 0.46^{**}$
	Root Part	Ethanol	$42.98 \pm 0.14^{**}$	$38.30 \pm 0.47^{**}$	$30.84 \pm 0.46^{*}$	$16.01 \pm 1.15^{**}$
		Water	$46.19 \pm 0.82^{**}$	$32.69 \pm 0.03^{**}$	$24.15 \pm 0.45^{**}$	$10.68 \pm 0.19^{**}$
Doxorubicin	b		7.48 ± 0.06	3.20 ± 1.01	2.78 ± 0.05	2.06 ± 0.12
DMSO (dim	ethyl sulfoxide) ^c		0	0	0	0

^a Values are expressed as $IC_{50} \pm SD$ (n=3).

^b Doxorubicin is used as a positive control.

^c DMSO; dimethyl sulfoxide, negative control.

**P value of <0.01 and *P value of <0.05.

Lactate Dehydrogenase (LDH) Activity Results

In this assay, the release of LDH from the A549, H1299 and C6 human cancer cells was measured after treatment with *A. elongatus* subsp. *nucleiferus* extracts. The leakage of LDH was measured in regards of IC_{50} values in a time dependent manner for 24, 48, and 72h treatment period. (Table 2). LDH release results of the extracts were presented in Table 2 for each cancer cells in a time dependently.

As presented in the Table 2, the IC₅₀ values for A549, H1299, and C6 cancer cells were calculated as comparing with non-tumor HUVECs and doxorubicin as positive control (IC₅₀= $2.48\pm0.02-3.02\pm0.46\,\mu$ g/mL). The extracts obtained from aerial parts and roots of the

plant were demonstrated the highest LDH activity towards H1299 cells, which was followed by A549 cells, and the results were found to have in accordance with anticancer activity results.

In another word, a significant increasing was observed in the IC₅₀ value from LDH by depending the time for all the tested extracts, which was supported by IC₅₀ values obtained from MTT assay. Among the plant part, root part was found to have high LDH activity (IC₅₀ values ranging from 12.64±1.01 to 38.11±0.64 µg/mL, for 24h), whilst, aerial part was found to have moderate LDH activity towards the cancer cells with IC₅₀ values in range of 27.82±1.02 – 51.83±1.21 µg/mL, after 24h treatment (Table 2).

Plant Part	Extract Type	Treatment		IC ₅₀ (µg/mL) values ^a		
		Hours	A549	H1299	C6	
Aerial Part	Ethanol	24 h	$30.16 \pm 0.18^{**}$	$27.82 \pm 1.02^{**}$	$51.83 \pm 1.21^{*}$	
		48 h	$49.85 \pm 0.83^{*}$	$35.06 \pm 0.68^{**}$	$76.05 \pm 0.03^{**}$	
		72 h	$67.42 \pm 0.12^{**}$	$65.16 \pm 0.62^{**}$	$78.46 \pm 1.01^{**}$	
	Water	24 h	$32.04 \pm 1.03^{**}$	$28.32 \pm 0.41^*$	$34.04 \pm 1.06^{**}$	
		48 h	$35.02 \pm 0.25^{**}$	$41.24 \pm 0.22^{**}$	$45.24 \pm 0.04^{**}$	
		72 h	$44.15 \pm 0.08^{*}$	$35.12 \pm 1.40^{*}$	$58.07 \pm 1.05^{**}$	
Root Part	Ethanol	24 h	$29.16 \pm 0.58^{**}$	$21.78 \pm 1.04^{*}$	$38.11 \pm 0.64^{**}$	
		48 h	$33.08 \pm 1.06^{**}$	$40.12 \pm 0.16^{**}$	$40.05 \pm 1.02^{**}$	
		72 h	$42.03 \pm 1.02^{**}$	$48.20 \pm 1.06^{**}$	$54.10 \pm 1.04^{**}$	
	Water	24 h	$21.38 \pm 0.78^{**}$	$12.64 \pm 1.01^{**}$	$29.06 \pm 0.80^{*}$	
		48 h	$26.48 \pm 0.12^{**}$	$18.06 \pm 0.09^{**}$	$36.07 \pm 1.64^{**}$	
		72 h	$35.10 \pm 0.26^{**}$	$24.06 \pm 0.34^{**}$	$42.01 \pm 1.04^{**}$	
Doxorubicin ^b			2.48 ± 0.02	2.54 ± 0.12	3.02 ± 0.46	

Table 2. LDH release results of A. elongatus subsp. nucleiferus e	extracts for A549, H1299, and C6 cells
---	--

 a The values are expressed as IC_{50} \pm SD (n=3) at 200 $\mu g/mL$ concentration.

^b Doxorubicin, positive control.

**P value of <0.01 and *P value of <0.05.

Releasing of LDH enzymes from the cells indicates damaging in the cells, since this enzyme releases from the necrotic cell membrane (Chan et. al., 2013). In view of this point, measuring the activity of LDH enzyme gives information about the percentage of dead and necrotic cells, as observed in this study

According to previous reports, there has been no study so far examining anticancer activity of *A. elongatus* subsp. *nucleiferus* extracts. On the other hand, this is the first evaluation of *in vitro* anticancer and antiproliferative activities of the extracts obtained aerial and root part of the plant by combining with cytotoxic activities of the cells and LDH enzyme activities that releases from the necrotic cells.

CONCLUSION

study, potential In current anticancer and antiproliferative activities of aerial part and root extracts obtained from A. elongatus subsp. nucleiferus were analyzed in detail by using various in vitro methods. Overall, the results revealed that the root extracts of the plant could have remarkable anticancer and antiproliferative activities through enhancement of apoptosis and necrosis, even at lower concentration and with a minimum exposure time. The data obtained from this studies could be considered as the first report for the literature, and the results should be validated by using further in vitro and in vivo techniques to assure of the molecular mechanisms underlying these wide ranges of anticancer effects of the plant.

ACKNOWLEDGMENTS

This study was financially supported by Scientific Research Project Unit of Kilis 7 Aralık University, Kilis-Turkey (Project code no: 1859LTP1). Authors would like to thank Gaziantep University, Medical Biology Department, Genetics Research Laboratory for their technical support.

Conflict of interests

No conflict of interest.

REFERENCES

- Al-Qubaisi M, Rozita R, Yeap SK, Omar AR, Ali AM 2011. Selective cytotoxicity of goniothalamin against hepatoblastoma HepG2 cells. Molecules 16(4): 2944-2959. https://doi.org/10.3390/ molecules 16042944.
- Block KI, Mead MN 2003. Immune system effects of echinacea, ginseng, and astragalus: a review. Integrative Cancer Therapies 2(3): 247-267. https://doi.org/10.1177/1534735403256419.
- Chan FK, Moriwaki K, De Rosa MJ 2013. Detection of necrosis by release of lactate dehydrogenase activity. Methods in Molecular Biology 979: 65-70. https://doi.org/10.1007/978-1-62703-290-2_7.
- Gezici S 2019a. Anticancer, antiproliferative, lysosomal and lactate dehydrogenase inhibitory effects of fruit extracts from sumac (*Rhus coriaria* L.) on human lung cancer cells. Acta Oncologica Turcica 52(1): 160-168. https://doi.org/10.5505/ aot.2019.09326.
- Gezici S 2019b. Comparative anticancer activity analysis of saffron extracts and a principle component, crocetin for prevention and treatment of human malignancies. Journal of Food Science and Technology 1-9. https://doi.org/10.1007/s13197-019-04014-y.
- Gezici S, Sekeroglu N 2019a. Current perspectives in the application of medicinal plants against cancer: novel therapeutic agents. Anticancer Agents in Medicinal Chemistry 19(1): 101-111. https://doi.org/10.2174/18715206196661812241210

04.

- Gezici S, Sekeroglu N 2019b. Neuroprotective potential and phytochemical composition of acorn fruits. Industrial Crops and Products 128: 13-17. https://doi.org/10.1016/j.indcrop.2018.10.082.
- Gezici S 2018. Promising anticancer activity of lavender (*Lavandula angustifolia* Mill.) essential oil through induction of both apoptosis and necrosis. Annals of Phytomedicine 7(2): 38-45. https://doi.org/10.21276/ap.2018.7.2.5.
- Gezici S, Serin Tanriöver C, Yayla F, Sekeroglu N 2018. Astragalus neurocarpus and Astragalus elongatus subsp. nucleiferus extracts as a Potential Source of Polyphenols with Antioxidant and Cholinesterase Inhibitory Activities. The 4th International Mediterranean Symposium on Medicinal and Aromatic Plants (MESMAP-4). April 18-22, 2018, Antalya, Turkey. Abstract Book, p: 41. (Oral Presentation). www.mesmap.org.
- Gezici S, Sekeroglu N, Kijjoa A 2017. In vitro Anticancer Activity and Antioxidant Properties of Essential Oils from *Populus alba* L. and *Rosmarinus officinalis* L. from South Eastern Anatolia of Turkey. Indian Journal of Pharmaceutical Education and Research 51(3): 498-503. https://doi.org/10.5530/ijper.51.3s.74.
- Ionkova I, Shkondrov A, Krasteva I, Ionkov T 2014. Recent progress in phytochemistry, pharmacology and biotechnology of Astragalus saponins. Phytochemistry Reviews 13(2): 343-374. http://dx.doi.org/10.1007/s11101-014-9347-3.
- Mosmann T 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation

and cytotoxicity assays. Journal of Immunological Methods 65(1-2): 55-63. https://doi.org/10.1016/0022-1759(83)90303-4.

- Newman DJ, Cragg GM 2016. Natural products as sources of new drugs from 1981 to 2014. Journal of Natural Products 79(3): 629-661. https://doi.org/10.1021/acs.jnatprod.5b01055.
- Ranjbar M, Mahmoudian B 2015. An overview on cytogenetics of the genus *Astragalus* subgenus *hypoglottis* (Fabaceae). Caryologia 68(2): 109-124. https://doi.org/10.1080/00087114.2015.1032571.
- Roleira FM, Varela CL, Costa SC, Tavares-da-Silva EJ 2018. Phenolic derivatives from medicinal herbs and plant extracts: anticancer effects and synthetic approaches to modulate biological activity. Studies in Natural Products Chemistry 57: 115-56. https://doi.org/10.1016/B978-0-444-64057-4.000041
- Shao BM, Xu W, Dai H, Tu P, Li Z, Gao XM 2004. A study on the immune receptors for polysaccharides from the roots of *Astragalus membranaceus*, a Chinese medicinal herb. Biochemical and Biophysical Research Communications 320(4): 1103 -1111.https://doi.org/10.1016/j.bbrc.2004.06.065.
- Strober W 2015. Trypan blue exclusion test of cell viability. Current Protocols in Immunology 111(1): A3.B.1-A3.B.3.

https://doi.org/10.1002/0471142735.ima03bs111.

Yalcın FN, Piacente S, Perrone A, Capasso A, Duman H, Calıs I 2012. Cycloartane glycosides from Astragalus stereocalyx Bornm. Phytochemistry 73: 119-126. https://doi.org/10.1016/j.phytochem. 2011.09.011.